Design and Application of LEF, a library of chemical fragments with different Local Environment of Fluorine

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Outline

- Fluorine in drug discovery
- ¹⁹F-NMR Screening
- Development of a new descriptor (F-FP-L)
- Design a library of chemical fragments with different Local Environment of Auorine (LEF)
- Predict ¹⁹F chemical shift of new molecules
- Analyse different protein environments of fluorine in protein-ligand crystal structures



Fluorine in Drug Discovery

- Conformational influences (binding affinity)
 - Proline conformation / Ar-OCF₃ vs Ar-OCH₃/ gauche effect / ...
- Protein-ligand interactions (binding affinity)
 - H-bond, lipophilic contacts, multipolar interactions
- Influences on physico-chemical properties (logP, pK_a)
- Bioavailability (absorption and transport)
- Metabolic stability (increased oxidative stability against enzymatic attack)
- ¹⁹F NMR Spectroscopy



Screening with ¹⁹F NMR Spectroscopy

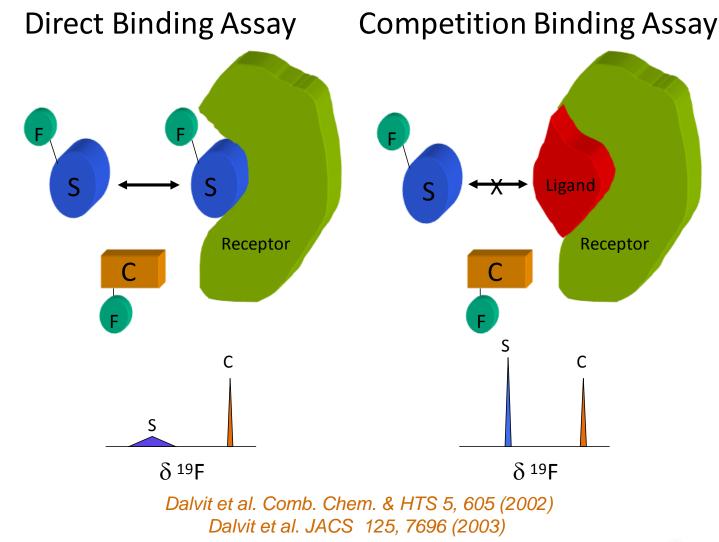


- Good sensitivity (0.83 ¹H)
- 100% Natural Abundance ¹⁹F NMR Active Isotope
- No problems with overlap
- No interferences from buffer, solvent, detergent signals
- ~ 15% of ACD molecules contain F
- Favourable Transverse Relaxation

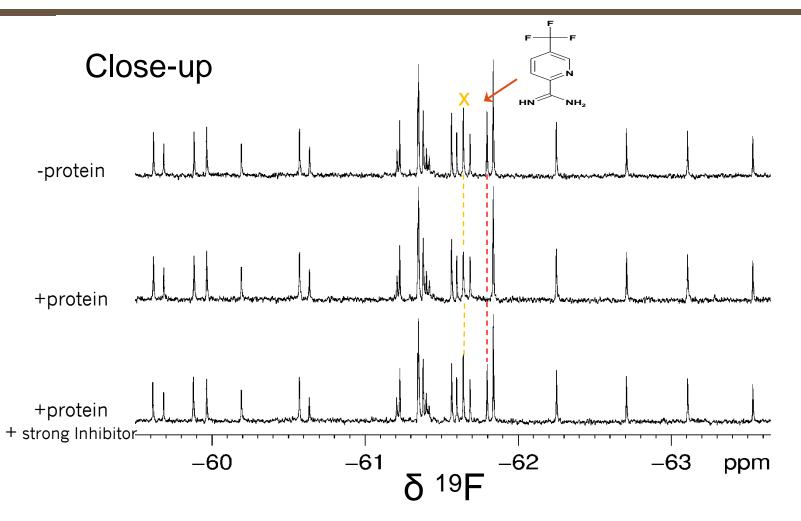
Dalvit C., Progress in NMR Spectroscopy 51, 243 (2007) Dalvit C., Concepts in Magnetic Resonance 32A, 341 (2008)



FAXS (Fluorine chemical shift Anisotropy and eXchange for Screening)



Screening with Large Mixtures (36 Molecules) + Validation of the Hits via Competition Binding Experiments



Vulpetti, A., Hommel, U., Landrum, G., Lewis, R., Dalvit, C., J. Am. Chem. Soc. 131 (2009) 12949-12959

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LEF (Library of Environment of Fluorine)

 Goal: design, construction and implementation of a diverse fragment library of fluorinated compounds for ¹⁹F NMRbased screening

Why Fluorinated fragments:

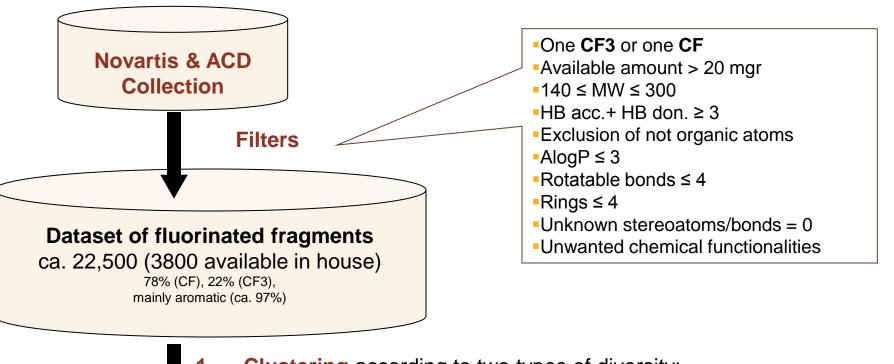
- Exploit high sensitivity ¹⁹F-NMR Screening technology
- Identification of spy molecules for subsequent screening of compounds containing (or not) fluorine atom(s)

• Why Fluorine in different local environment:

 Fluorine Local Environment is relevant for the protein-fluorine interactions. Identification of fluorophilic hot-spots by presenting the fluorine with different chemical and structural features.



Key steps

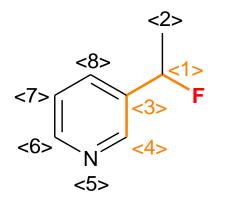


- 1. Clustering according to two types of diversity: <u>global</u> (on the whole molecule) and <u>local</u> (based on the description of the 2D local chemical environment around the F atom and the CF_3 group).
- 2. Annotation of other calculated properties (synthetic tractability, in silico water solubility)
- 3. Selection by visual inspection

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Fluorine local-environment fingerprints (F-FP-5)

- New descriptor derived from the topological-torsion descriptor concept
- Paths consisting of between one and five bonds and only starting from the fluorine atom or CF3 moiety are included



E.g. highlighted path: F-<1>-<3>-<4>

For each atom: Element, Number of π electrons, Number of heavy neighbors (not counting those in the torsion).

F-0-0,C-0-1,C-1-1,C-1-1

Vulpetti, A., Hommel, U., Landrum, G., Lewis, R., Dalvit, C., J. Am. Chem. Soc. 131 (2009) 12949-12959

Fluorine local-environment fingerprints (F-FP-5)

The set of all paths of length one to five bonds rooted at the fluorine atom are enumerated

	Length	Path	Atom Codes	Bit Id
	<2> 1	F-C1	(F-0-0,C-0-2)	b1
•	2	F-C1-C2	(F-0-0,C-0-1,C-0-0)	b2
<8>	<1> 2	F-C1-C3	(F-0-0,C-0-1,C-1-2)	b3
<7>	F 3	F-C1-C3-C4	(F-0-0,C-0-1,C-1-1,C-1-1)	b4
	<3> 3	F-C1-C3-C8	(F-0-0,C-0-1,C-1-1,C-1-1)	b4
<6>	<4>	F-C1-C3-C4-N5	(F-0-0,C-0-1,C-1-1,C-1-0,N-1-1)	b5
	4	F-C1-C3-C8-C7	(F-0-0,C-0-1,C-1-1,C-1-0,C-1-1)	b6
<5>	5	F-C1-C3-C4-N5-C6	(F-0-0,C-0-1,C-1-1,C-1-0,N-1-0,C-1-1)	b7
	5	F-C1-C3-C8-C7-C6	(F-0-0,C-0-1,C-1-1,C-1-0,C-1-0,C-1-1)	b8
Similarity	$Sim(FP_i, FP_j) = \frac{2 \sum_{b} \min(FP_{ib}, FP_{jb})}{\sum_{b} FP_{ib} + \sum_{b} FP_{jb}}$	_)		10 = 0.8

Vulpetti, A., Hommel, U., Landrum, G., Lewis, R., Dalvit, C., J. Am. Chem. Soc. 131 (2009) 12949-12959

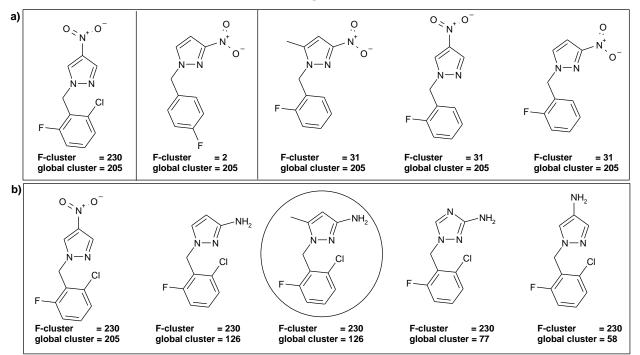
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Clustering

Based on:

- local fluorine-environment fingerprints (F-FP-5),
- whole molecular structure (global description, FCFP-4).

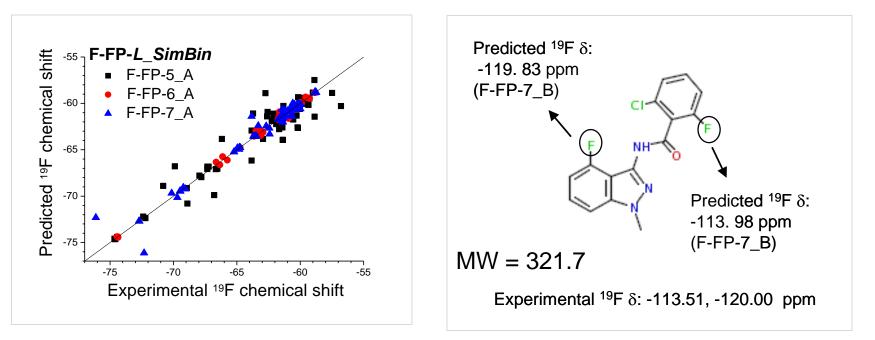


Molecules of the same F-cluster have similar ¹⁹F chemical shifts



Extension of F-FP-L descriptor to ¹⁹F chem. shift prediction

 New extended fluorine fingerprint and a distance-weighted k-nearest neighbors algorithm applied on a training set of known chemical shifts.



The quality of the prediction depends on both the training set size and its diversity. The expansion of our current chemical shift training set is on going.

Vulpetti, A., Landrum, G., Rüdisser, S., Erbel, P., Dalvit, C., J. of Fluorine Chem., 131 (2010) 570-577

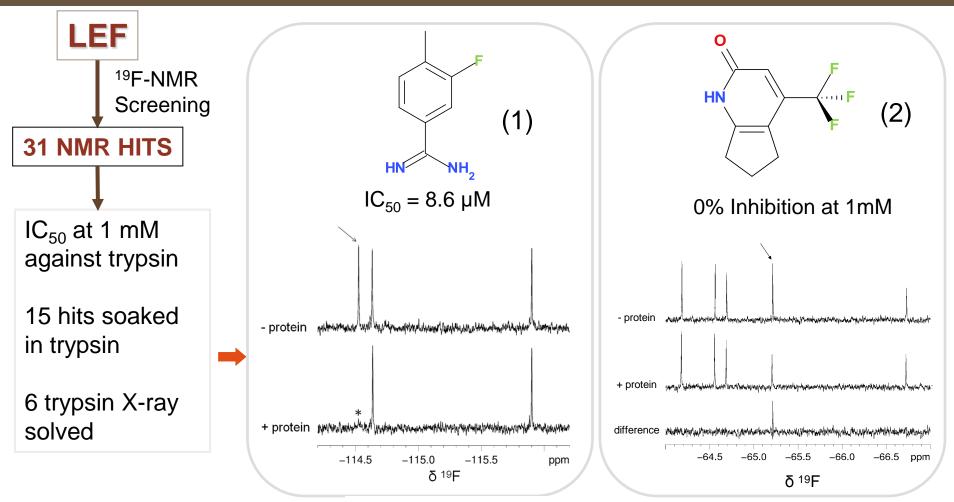
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Experimental Conditions

- Current LEF: 120 mixtures (54 CF₃ and 66 CF) (12 molecules per mixture) (expansion in progress)
- Mixture concentration : CF₃ and CF mixtures are tested at 15-20 μM and 35-40 μM, respectively with 600 MHz without cryoprobe and ~ 5 μM and ~ 15 μM, respectively with 600 MHz with cryoprobe
- Protein concentration : 1-10 μM
- Acquisition time for each mixture is 10-15 minutes [one 96-wells rack (1152 molecules) in < 24 hours]
- Spectral analysis performed with peak peaking
- Tested with success on different targets



LEF Screening in Trypsin



Vulpetti A., Schiering N., Dalvit C. Proteins: Structure, Function, and Bioinformatic Journal (2010) - DOI: 10.1002/prot.22836



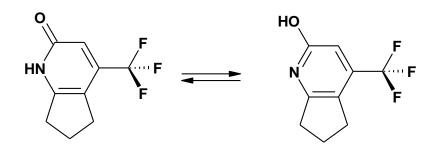
His57 Ser21 Ser195 Gln192 Asp189

Compound (1) $IC_{50} = 8.6 \,\mu M$

- Compound (1) is modeled in one major and two minor orientations of the ring with occupancies of 54% (orientation-1), 22% (orientation-2) and 24% (orientation-3)
- Orientation-1 (orange):
 - the fluorine points towards Ser195 (OG and NH)
- Orientation-2 (*white*):
 - the aromatic ring is flipped by ca. 180° and the fluorine is pointing in the opposite direction, at a distance of ca. 3 Å to the Gln192 side chain NE2.
- Orientation-3 (yellow):
 - At 3.6 Å to Ser195 OG atom, at 3.2 Å to the carbon of the carbonyl of Ser214, $(\alpha_1 = C-F...C \text{ of } 115.5^\circ; \text{ angle } \alpha_2 = F...C=O \text{ of } 69.9^\circ$ (favorable multipolar interaction)

Compound (2)

Compound (2) can be drawn in two tautomeric forms: 2-pyridone and 2-hydroxypyridine.



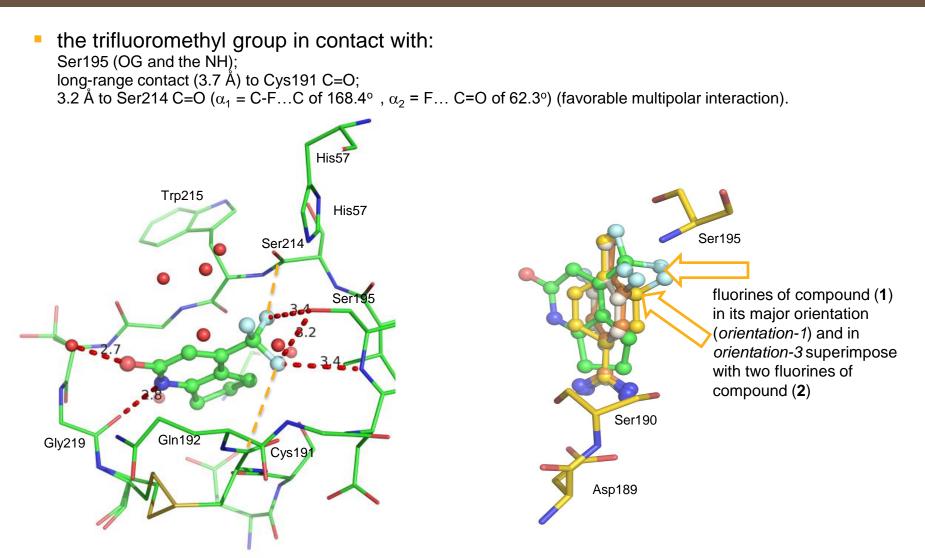
• The 2-pyridone tautomeric form is estimated to be in general the preferred tautomer considering the occurrences of the two tautomeric forms in CSD and PDB databases.

Name	∆G (kcal mol ⁻¹) /	Major form in Water (M)		Minor form in Water (m)			PDB with	
	medium / relevant form(s)	Structure/Name	CSD	PDB	Structure/Name	CSD	PDB	identical HB score
2-pyridone	4.2/water/M; -0.65/gas (calc.)/m; ND/solid/m.		172	24		12	2	2

Milletti F., Vulpetti A. J.Chem.Inf.Model., 50 (2010) 1062-1074



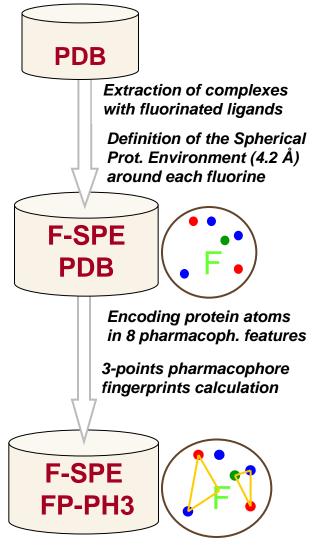
Compound (2) 0% Inhibition at 1 mM



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F-SPE-PDB database

preparation



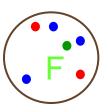
Protein sub-sites are extracted around each fluorine (F-SPE) contained in the PDB and compared to the query of interest by using a pharmacophoric description

Feature	Description	Description
D	donor attribute	NZ.LYS
CA	alpha carbon	CA
C=	carbon of carbonyl of	C, CG.ASP, CG.ASN, CD.GLU, CD.GLN,
	amides / C guanidinium	CZ.ARG
Р	donor and acceptor (-	OG.SER, OG1.THR, OH.TYR, ND1.HIS,
	OH)	NE2.HIS
Н	hydrophobe (-CH ₂ ,-CH ₃)	all aliphatic CB, CG, CD and SD.MET, SG.CYS
D=	π donor (e.g., sp ² NH)	N, NE2.GLN, ND2.ASN, NH1.ARG, NH2.ARG,
		NE.ARG, NE1.TRP
A=	π acceptor (e.g., sp ² O)	O, OD1.ASP, OD2.ASP, OE1.GLU, OE2.GLU
H=	π hydrophobe	all aromatic carbons of Phe, Tyr, Trp, His
X	Ligand fluorine atom	F

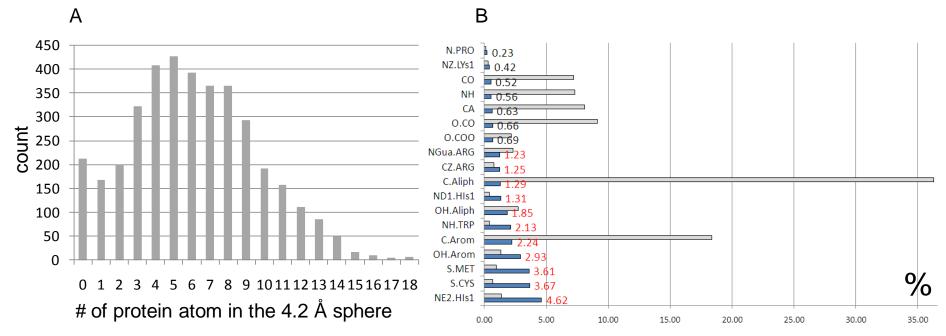
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F-SPE-PDB database

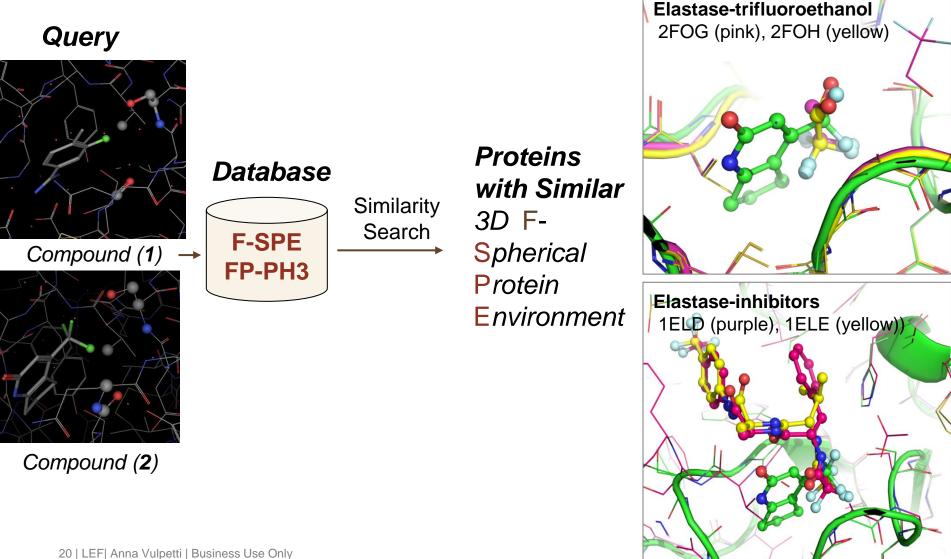


- A. Number of complexes with 0 to 18 **protein atoms** in the sphere of radius 4.2 Å. [Ave= 6.14, Std= 3.49]
- B. Frequency / Relative Propensity of the 18 protein atom types



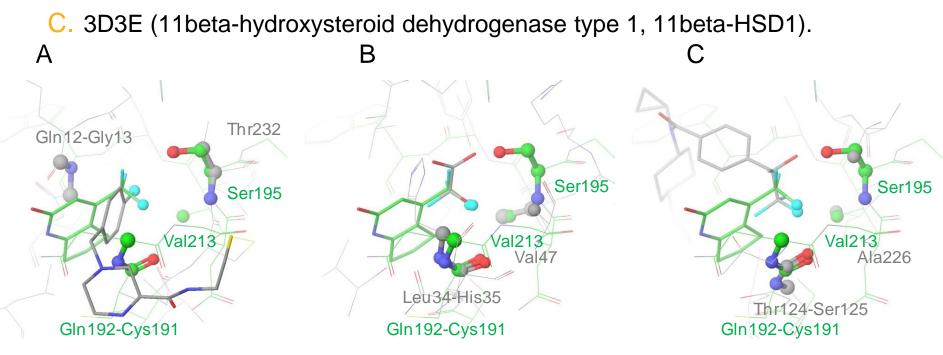
- The **number of triplets** is on average 39 with a standard deviation of 13
- Clustering : 1543, 2177 and 2414 clusters (Tanimoto coefficient 0.6, 0.7, 0.8)

Overlay of Compound (2) bound to Trypsin with TFE and Inhibitors bound to Elastase



Overlay of Compound (2) bound to Trypsin with Ligands bound to different Proteins

- A. 2ZJJ (BACE1),
- B. 3CEE (oxylipin-conjugated barley lipid transferase protein, LTP1) and



Despite their different global fold and their different active binding sites these proteins have an environment around the ligand fluorine atoms that is quite similar

Summary

¹⁹F-NMR screening of LEF

- Identification of useful starting fragments for fragment-based drug discovery projects
- Characterization of Fluorophilic Protein Environments (synergistic combination of ¹⁹F-NMR, X-Ray, CADD)
 - Generation of the F-SPE-PDB database
 - Similarity searches into the F-SPE-PDB database
 - Identify proteins accommodating fluorine atoms with a similar pattern recognition.
 - Virtual F-scan.



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