Automated SPR Data Processing and Sensogram Classification

Constantine Kreatsoulas Chemistry Modeling & Informatics – West Point, PA Merck & Co., Inc. constantine_kreatsoulas@merck.com

Where patients come first 📀 MERCK



Abstract

MERCK

Fragment-based approaches have added to the arsenal of tools used to identify novel small molecule leads with high ligand efficiencies. A variety of label-free technologies have been developed and implemented throughout the industry for fragment screening. Using surface plasmon resonance (SPR) as a fragment screening platform is in its infancy. The miniaturization and automation of this technology has led to the associated problem of dealing with the large volume of raw data generated. The resources needed for the analysis, integration and prioritization of compounds screened makes it challenging to take the results of an SPR screen into the workflow of project teams engaged in the discovery process in a timely fashion. As such, several sets of equations were derived and implemented on Merck's intranet to score single sensograms to distinguish stable binders from weak or anomalous binders. This set of equations was optimized and validated on simulated data to both capture "fragment-like" behavior from SPR experiments and to filter out much of the anomalous behavior commonly observed. This talk will cover the creation of the theoretical data sets, the resultant equations, and the performance metrics used to evaluate them.

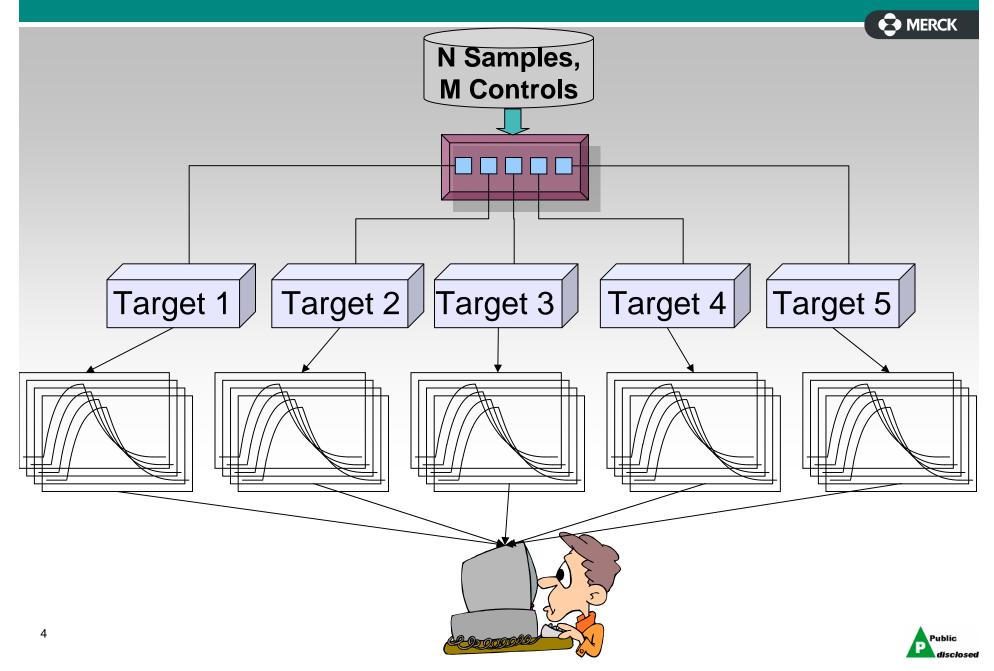


Discovery Project Needs Drove Collaboration

- Fragment Screening in the Discovery Process
 - Lead ID time is critical!
 - Need rapid turnaround of results
 - Assay development takes time for novel targets
 - Biacore A100 assay takes little time (N fragments + M controls on 5 spots)
 - Visual analysis of each of the $5 \times (N + M)$ curves
 - ~1 week (Bottleneck)
- Selection criteria for follow up
 - Response relative to theoretical R_{max}
 - Appropriate curve shape (box) for fragment-like kinetics
 - Selectivity over other targets



The Problem in a Nutshell



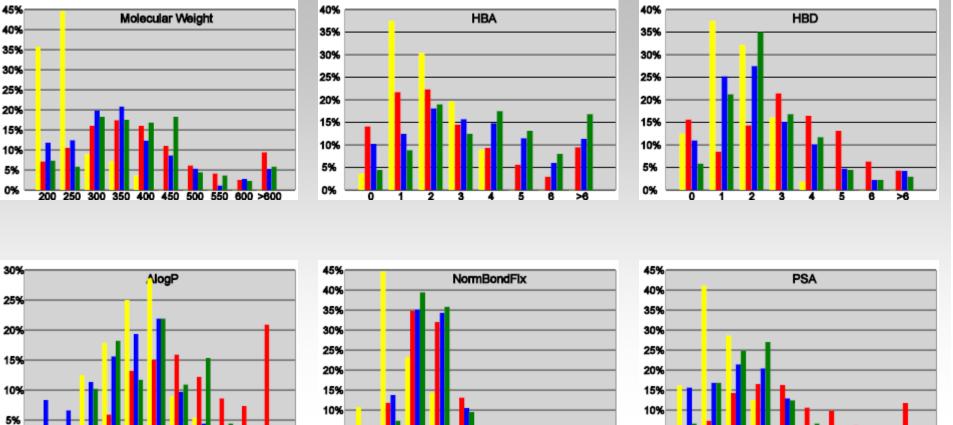
The Problem in a Nutshell

How can we speed this up to have an immediate project impact?





Aside: Summary of Merck Fragment Library Properties



Yellow: Fragments Assayed Red: All Launched Drugs as of 2006 (MDDR) Blue: Orally Delivered Drugs Green: Top 200 Small Molecule Drugs of 2008

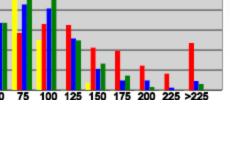
0.6 0.7 0.8 >.8

5%

0%

0

0.1 0.2 0.3 0.4 0.5

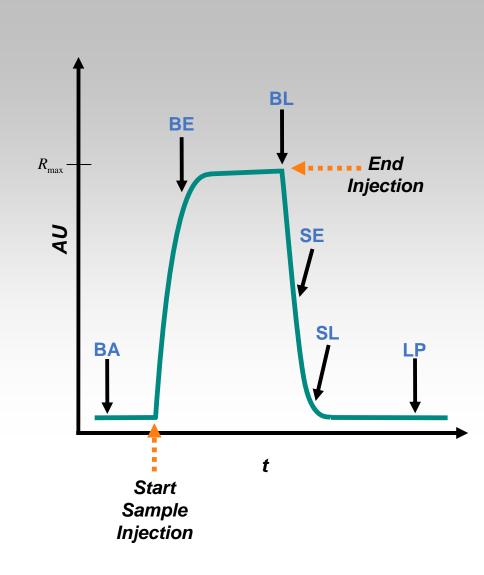


5%

0%

Public

Another Aside: A Single Sensogram



Single-point experiment

- Not titrated
- > 1500 compounds/screen

$$R_{\max} = \frac{n \cdot R_L \cdot MW_{analyte}}{MW_{ligand}}$$

Reporter Points

- BA: baseline (15 sec window)
- BE: binding early (10 sec)
- BL: binding late (10 sec)
- SE: stability early (10 sec)
- SL: stability late (10 sec)
- LP: late points (10 sec)

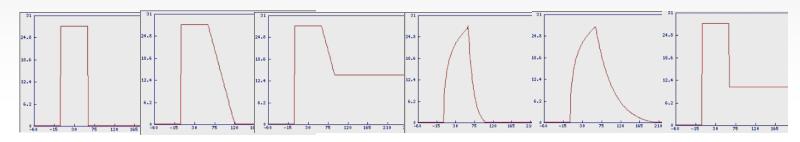
Theory vs. Experiment 🔁 MERCK Many curve types, need to account for them all: The Good 75 51 58 38.8 45.2 41 26.6 31.4 24 14.4 17.6 ma 2.2 3.8 -14.125 10.75 35.625 60.5 85.375 110.25 135.125 -14.125 10.75 35.625 60.5 85.375 110.25 135.125 85.75 110.5 135.2 -13.25 11.5 36.25 61 -11 The Bad and Ugly 39.6 48.4 41.2 27.2 33.8 28.4 14.8 19.2 15.6 2.4 4.4 2.8 85.75 110.5 135.25 -13.2511.5 36.25 61 39 14.125 10.75 35.625 60.5 85.375 110.25 135.125 -13.25 11.5 85.75 110.5 135.25 36.25 61



Theoretical Data Sets

Used to mimic experimental data and benchmark ranking equations

Data Set	IS1	IS2	IS3	IS4	IS5	IS6
Design Criteria	Fast-on, fast-off kinetics (fragment- like)	Fast-on, slow- off kinetics	Fast-on, slow-off; incomplete dissociation	Slow-on, moderately fast-off (high affinity)	Slow-on, slow-off kinetics (drug- like)	Fast-on,fast- off kinetics; incomplete dissociation



24 total "training" sets:

- Curves modulated from 100%, 75%, 50% and 25% of R_{max} .
- Two modulated to 150% of R_{max}.



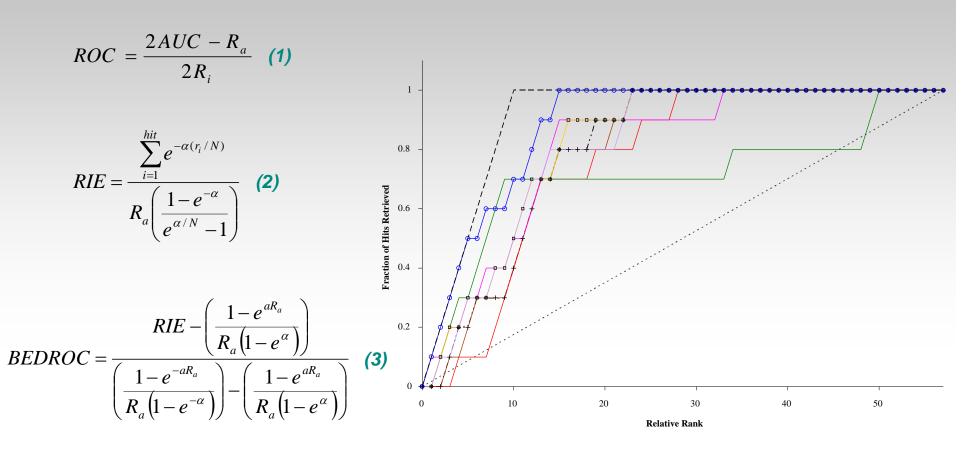
How to Evaluate the Resultant Rankings?

Mathematically:

AUC = Area Under Accumulation Curve

Accumulation Curve

Graphically:





Possible Ranking Metrics

Simple, widely used: $S = \frac{B_L - A_L}{R_L}$

$$S = \frac{B_L - B_A}{R_{\text{max}}}$$
 (4) $S_{Occ} = 100 \cdot \frac{B_L}{R_{\text{max}}}$ (5)

Penalize slow on-rate:

$$S = \frac{2B_{L} - (B_{E} + B_{A})}{R_{\text{max}}}$$
 (6)

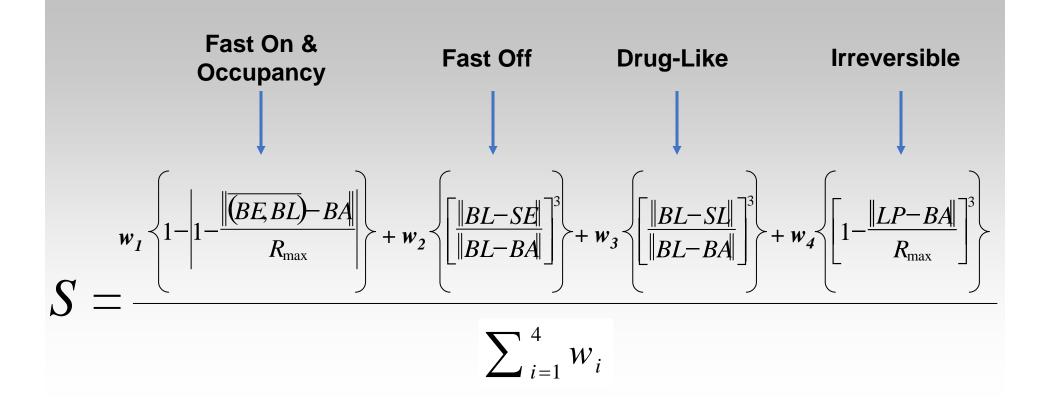
$$S = \frac{B_L - S_E}{R_{Max}}$$
 (7) $S = \frac{B_L - (2S_E - S_L)}{R_{max}}$ (8)

Penalize aggregators:

$$S = \frac{B_L - L_P}{R_{Max}}$$
 (9) $S = \frac{B_L - (2S_L - L_P)}{R_{max}}$ (10)

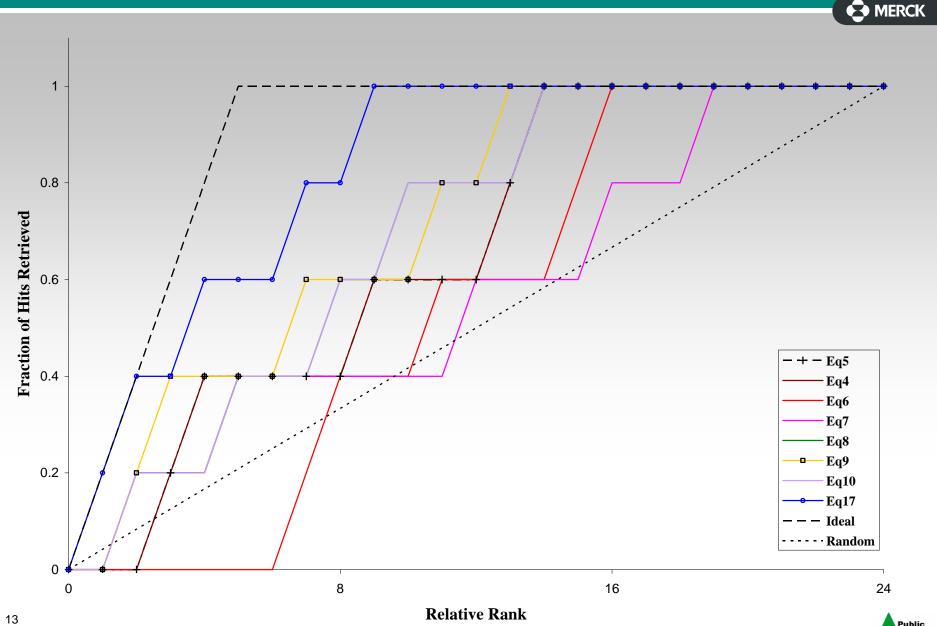
or combine them...

Combined Ranking Metric (aka Eq. 17)



Weights, w_i , allow for a single equation to be used for fragment and small molecule ranking.

Performance on Theoretical Data Set



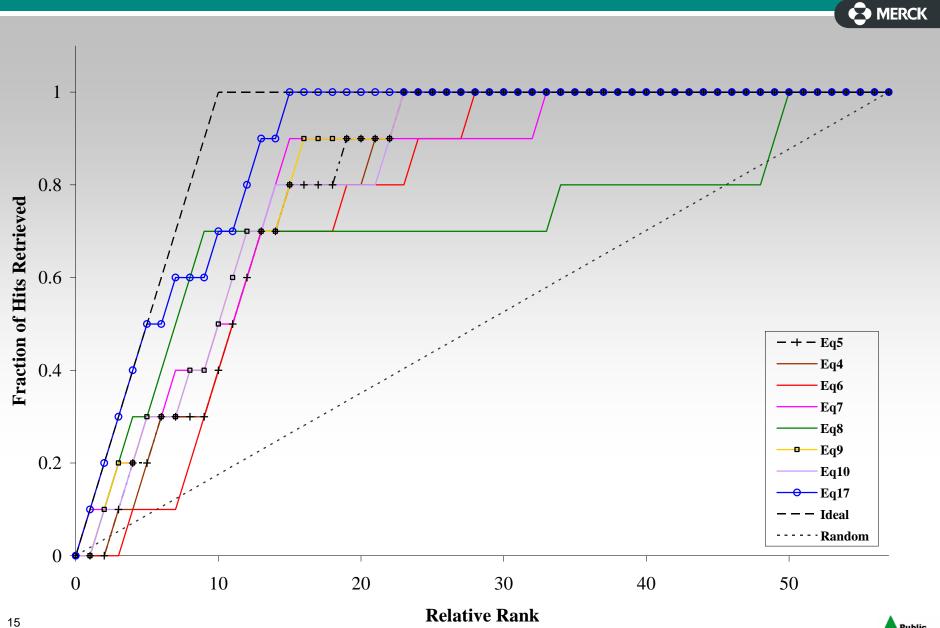
Public P disclosed

Performance on Theoretical Data Set

	AUC	ROC	RIE	BEDROC
Eq. (4)	0.65	0.71	2.34	0.49
<i>Eq.</i> (5)	0.65	0.71	2.34	0.49
Eq. (6)	0.53	0.56	1.15	0.24
Eq. (7)	0.55	0.59	1.93	0.41
Eq. (8)	0.68	0.76	2.36	0.50
Eq. (9)	0.71	0.79	2.76	0.58
Eq. (10)	0.68	0.76	2.36	0.50
Eq. (17)	0.82	0.93	3.55	0.75
Ideal	0.89	1.02	4.48	0.95
Random	0.49	0.52	1.76	0.37



Performance on a Fragment Screen



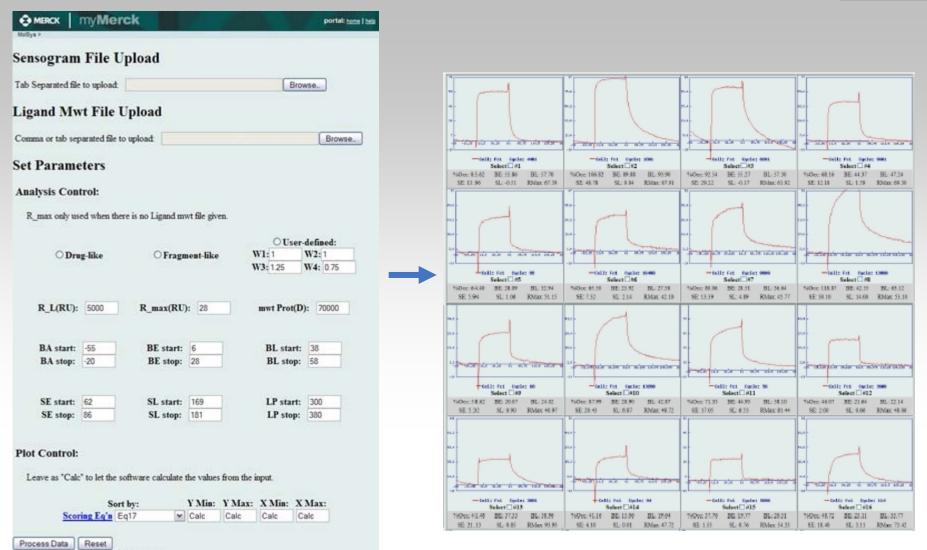


Performance on a Fragment Screen

	AUC	ROC	RIE	BEDROC
Eq. (4)	0.80	0.87	1.91	0.35
Eq. (5)	0.81	0.87	1.92	0.35
Eq. (6)	0.78	0.83	1.13	0.20
Eq. (7)	0.81	0.88	3.31	0.60
Eq. (8)	0.81	0.88	2.51	0.45
Eq. (9)	0.82	0.89	2.76	0.50
Eq. (10)	0.81	0.88	2.51	0.45
Eq. (17)	0.89	0.97	5.18	0.94
Ideal	0.91	1.00	5.53	1.00
Random	0.49	0.49	1.85	0.33



Deployed via Intranet: Modify on the Fly



If nothing happens, press the button again.

Comments

Impact

- Shortened SPR screen analysis time > 50x
- Rapid selection of true positives
- Outperforms usual methods of fragment selection
- Can use for fragments or fully elaborated molecules

Next Steps

- Use of machine learning to select best w_i from a series of screens and users' rankings
- Optimization of w_i for drug-like molecules
- Would love to hear how this works in others' hands
- Not meant to replace titrations!

Acknowledgements

Automated Biotechnology

- Kartik Narayan
- Delphine Collin
- Krista Getty

Chemistry Modeling & Informatics

- Chris Culberson
- John Sanders

Thank you!

Reference:

Kreatsoulas & Narayan, Analytical Biochemistry, 402 (2010) 179-184.

