

Targeting Protein-Protein Interactions via Target-Guided Synthesis

Roman Manetsch



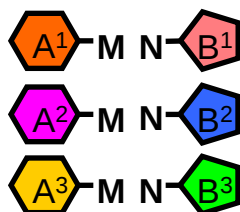
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Kinetic Target-Guided Synthesis

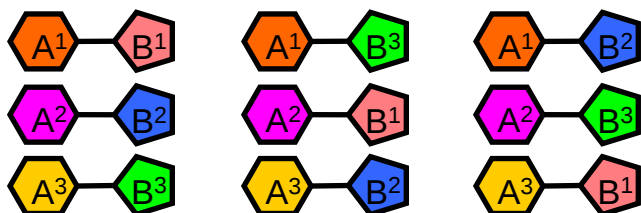
Fundamental Principle: Polyvalent interactions can be collectively much stronger than corresponding monovalent interactions.

Conventional Lead Discovery

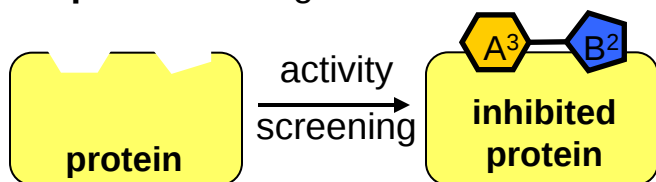
Step 1: Synthesis of building blocks



Step 2: Preparation of library of final compounds

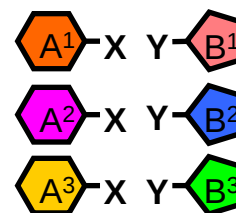


Step 3: Screening for inhibition

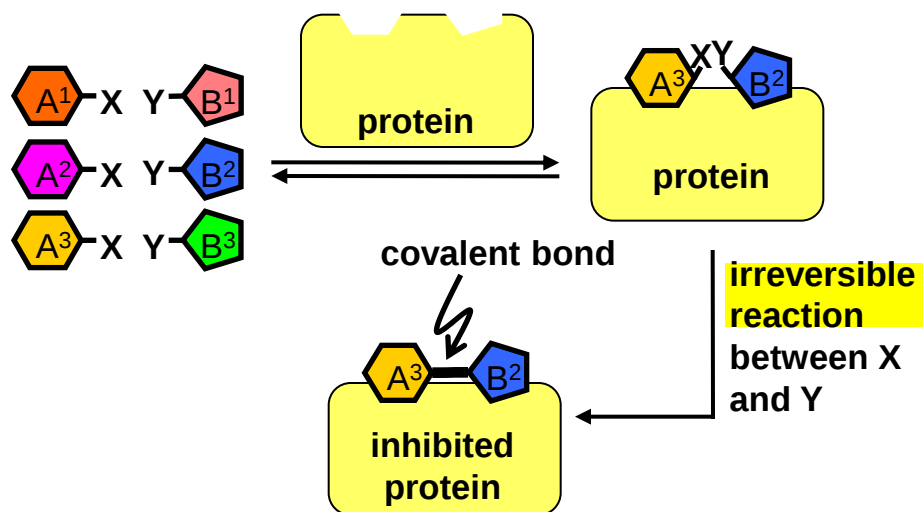


Kinetic Target-Guided Synthesis

Step 1: Synthesis of building blocks



Step 2: Screening and synthesis of biologically active compounds



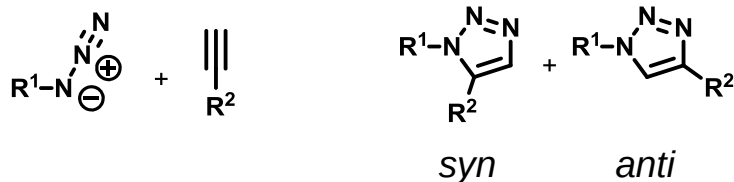
In Situ Click Chemistry – A Kinetic TGS Variant

Polyvalent interactions can be collectively much stronger than corresponding monovalent interactions.

Examples:

- hydrazone formation
- disulfide bond formation
- epoxide ring-opening
- N-alkylation
- S-alkylation

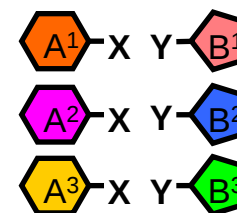
In situ Click Chemistry:



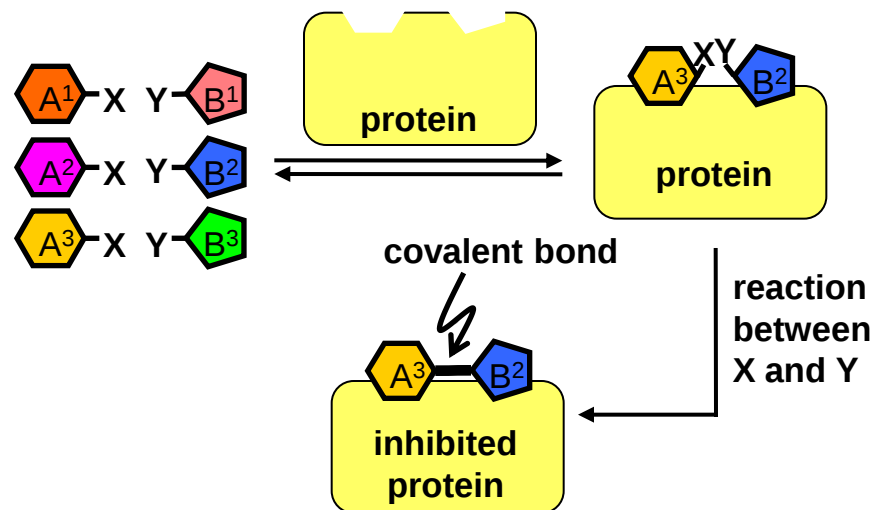
- modular
- wide in scope
- high-yielding
- insensitive to oxygen and water
- enzymatic targets
- potent inhibitors

Target-Guided Synthesis

Step 1: Synthesis of building blocks



Step 2: Screening and synthesis of biologically active compounds



In Situ Click Chemistry: Kinetic TGS on Enzymatic Targets



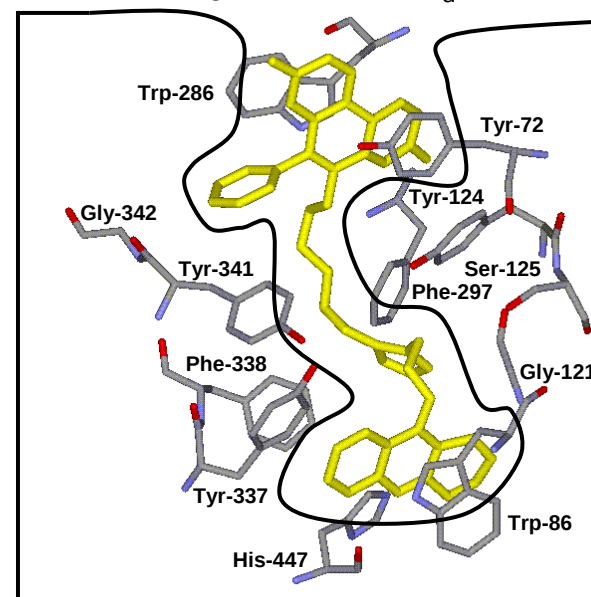
- good hit rate – known inhibitors as reactive building blocks
- very potent hit compounds
- reactive building blocks are binding with good/high affinity

for mouse AChE

TZ2 $K_d = 23$ nM

PA6 $K_d = 360$ nM

***syn*-TZ2PA6** $K_d = 410$ fM



- good hit rate – known inhibitors as reactive building blocks
- very potent hit compounds
- reactive building blocks are binding with good/high affinity

Questions

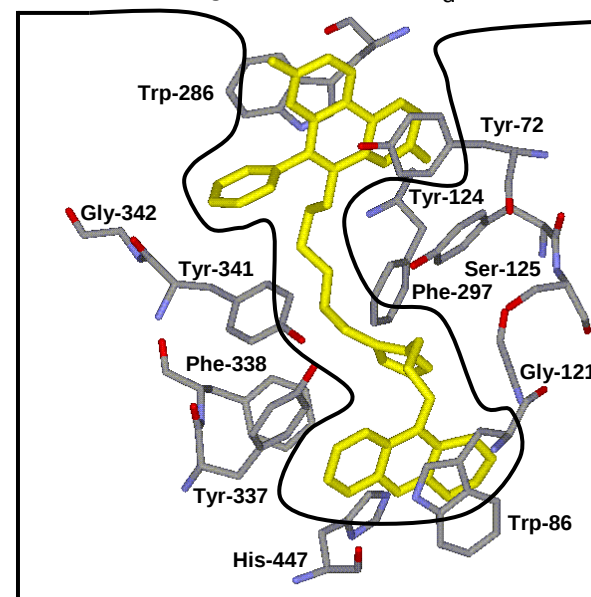
- Is kinetic TGS suitable for fragment linking?
- Is kinetic TGS suitable for challenging targets? Protein-protein interactions?
- Has kinetic TGS potential to be a straightforward fragment-based approach for targets limited in structural information?

for mouse AChE

TZ2 $K_d = 23$ nM

PA6 $K_d = 360$ nM

syn-TZ2PA6 $K_d = 410$ fM



Disrupting Protein-Protein Interactions

Theory of hot spots:

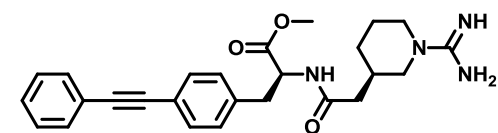
Subregions of protein-protein interfaces that contribute significantly to the overall free energy of binding between the proteins

Size of hot spots is comparable to the surface area of drug-like molecules

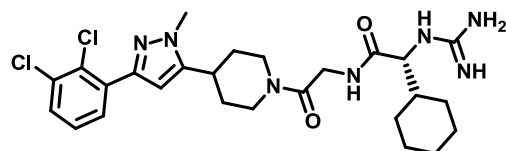
Flexible amino acid residues at the protein surface

Example: Interleukin-2 (IL-2) and its receptor IL-2 α R

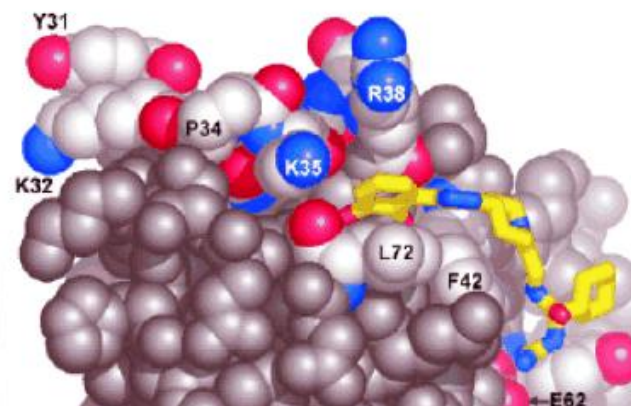
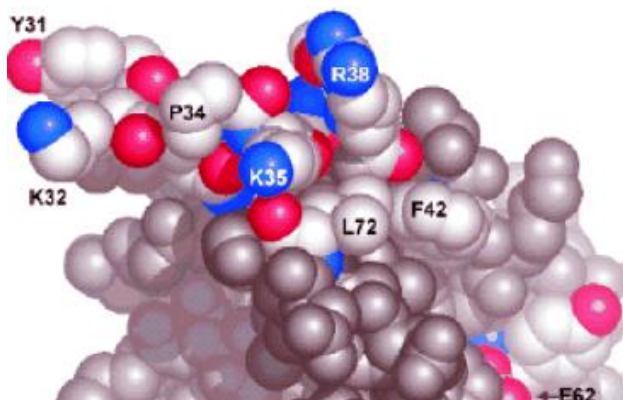
Compounds bind to the IL-2 α R-binding region on IL-2



$IC_{50} = 3 \mu M$



$IC_{50} = 6 \mu M$



The Bcl-2 Family: Apoptotic Switches in Cancer

Bcl-2 Family Proteins

- anti-apoptotic Bcl-2 family proteins: Bcl-2, Bcl-X_L and Mcl-1
- pro-apoptotic proteins: Bax, Bak and BH3-only proteins (Bim, Bad, Bid, Bik)

Category 1: anti-apoptotic

Bcl-2, Bcl-X_L, Bcl-w, Mcl-1, A1



Category 2: multidomain pro-apoptotic

Bax, Bak



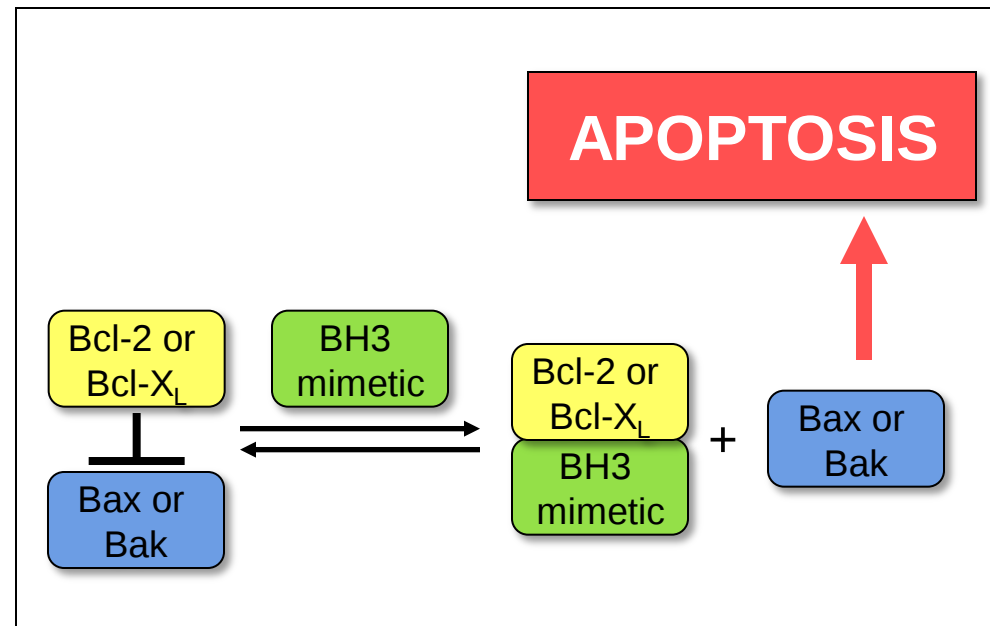
Category 3: BH3-only pro-apoptotic

Bad, Bid, Bim, Noxa, Puma

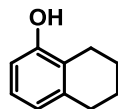


Bcl-2 Cancer Biology

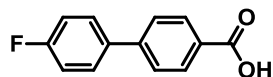
- central regulators of apoptosis (validated targets)
- contribute to tumor initiation, progression and resistance to therapy
- disrupting heterodimerization of anti- and pro-apoptotic proteins inhibits anti-apoptotic function of Bcl-2, Bcl-X_L, and Mcl-1



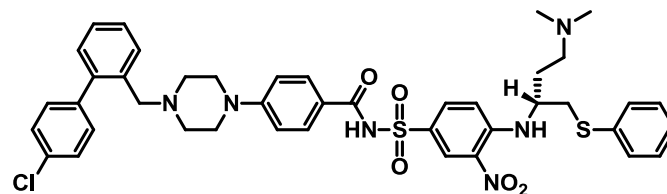
ABT-737 and Analogues



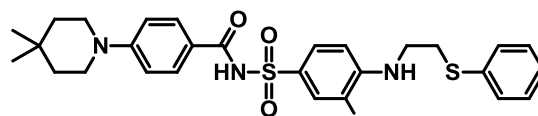
$K_D = 4.3 \text{ mM}$



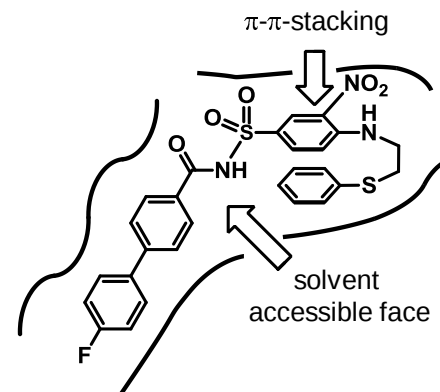
$K_D = 300 \text{ }\mu\text{M}$



ABT-737: $K_i < 2.0 \text{ nM}$

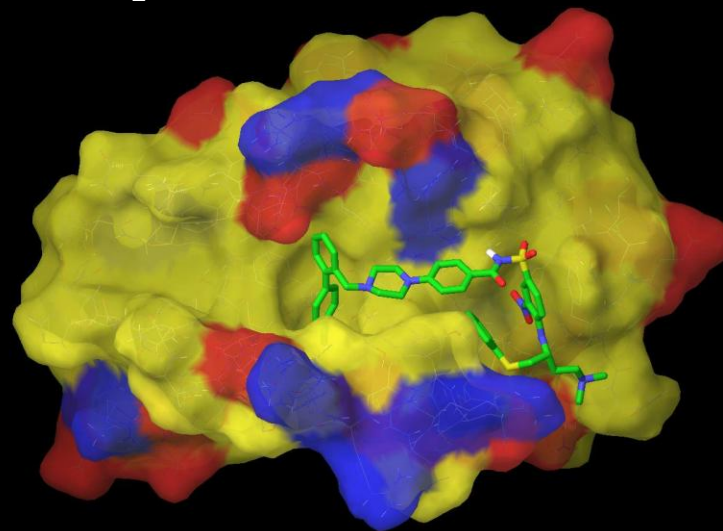


$K_i = 19 \text{ nM}$

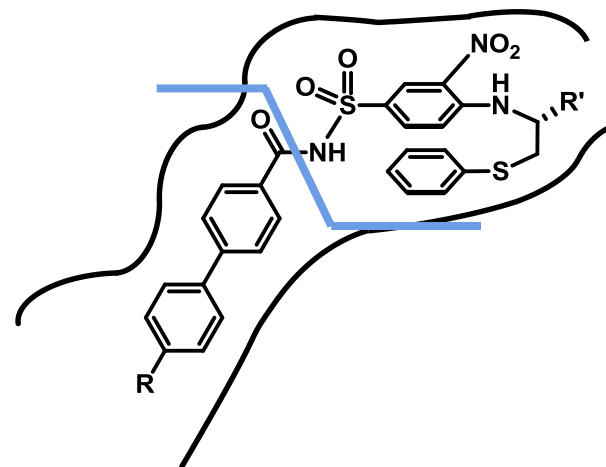
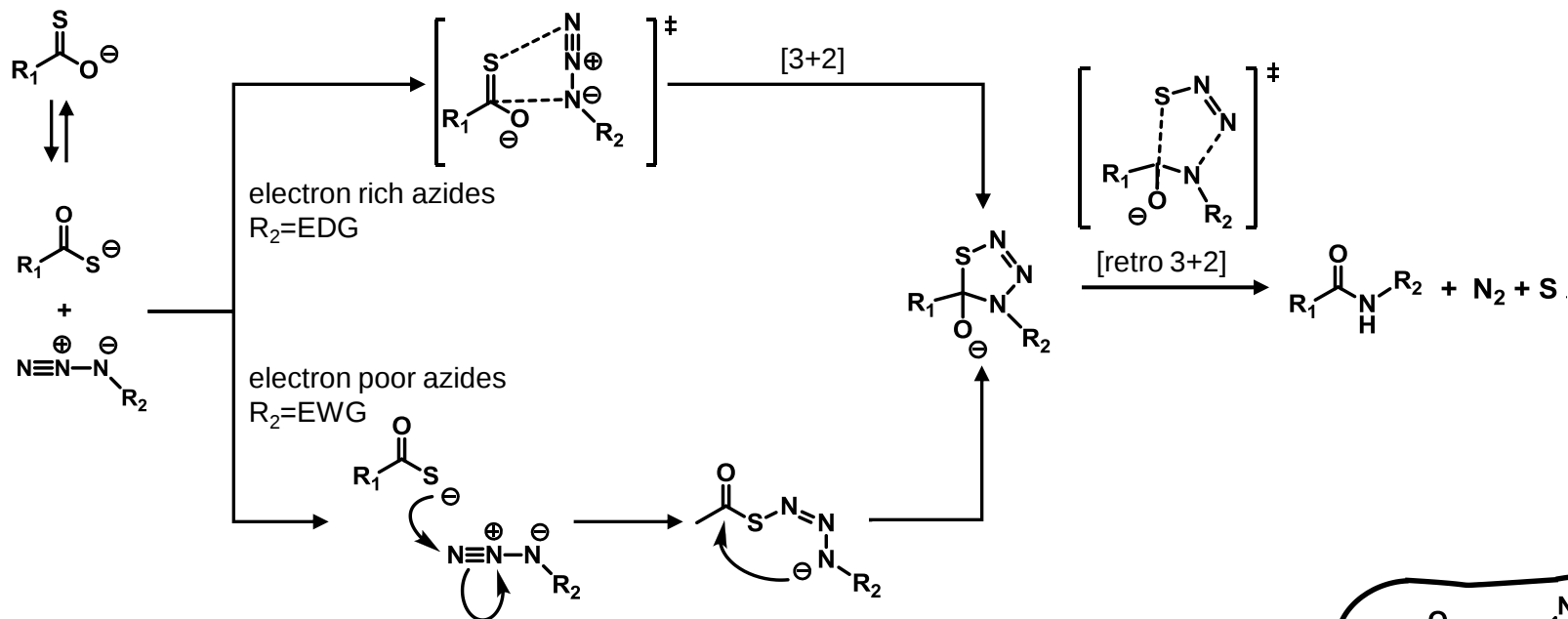


$K_i = 36 \text{ nM}$

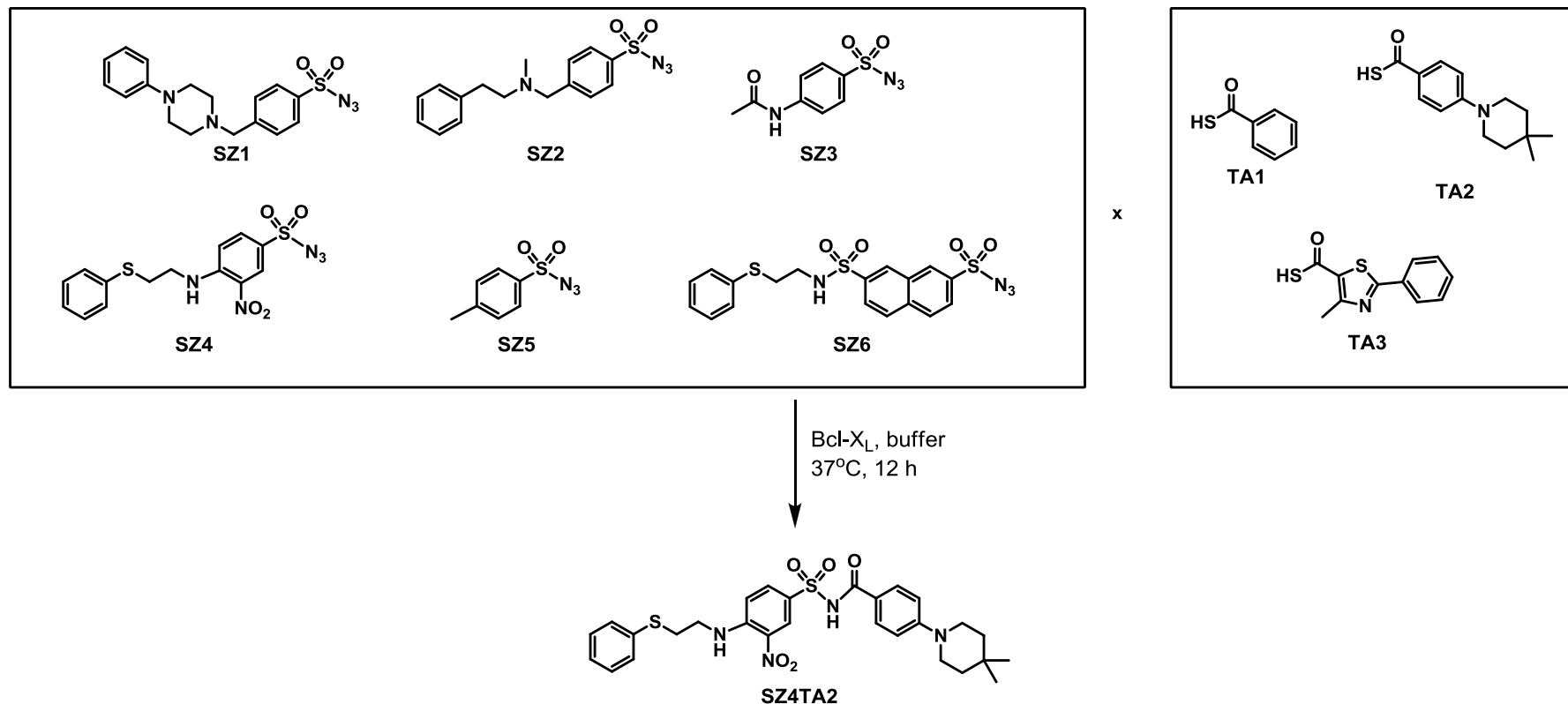
Bcl-X_L-ABT-737



Amidation Reaction



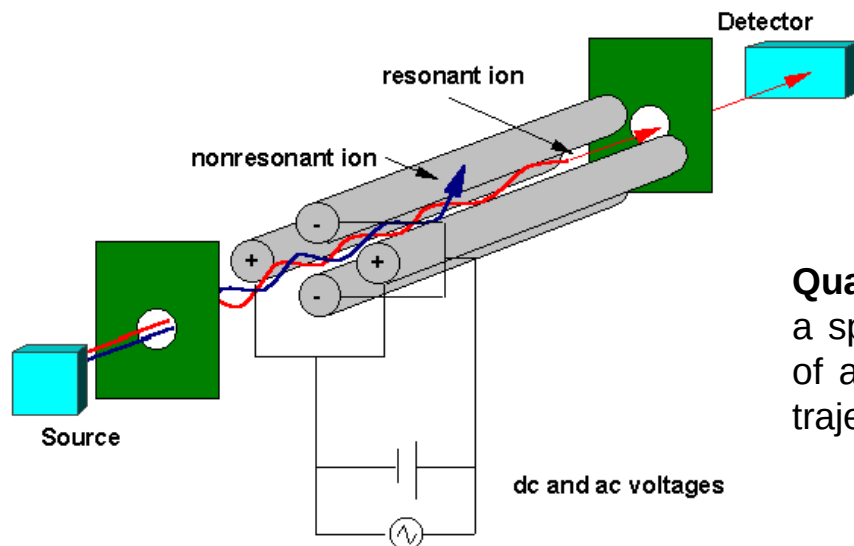
Proof of Concept: Bcl-X_L-templated Reactions



Incubation conditions: 2 μ M Bcl-X_L (phosphate buffer, pH=7.4)
 20 μ M sulfonfylazide and 20 μ M thioacid building blocks
 binary mixtures, 37°C, 12 h
 LC/MS-SIM analysis

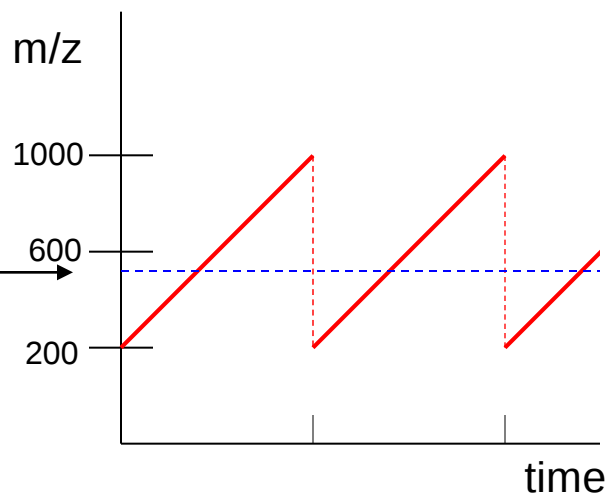
9 building block reagents $\rightarrow$ 18 combinations $\rightarrow$ 1 TGS hit compound

Selected Ion Monitoring *versus* Scan Mode

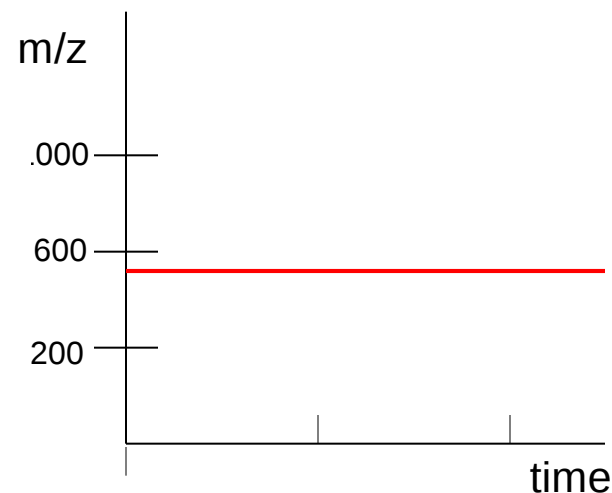


Quadrupole Mass Analyzer: at a specific value of voltage, ions of a particular m/z follow stable trajectory to detector.

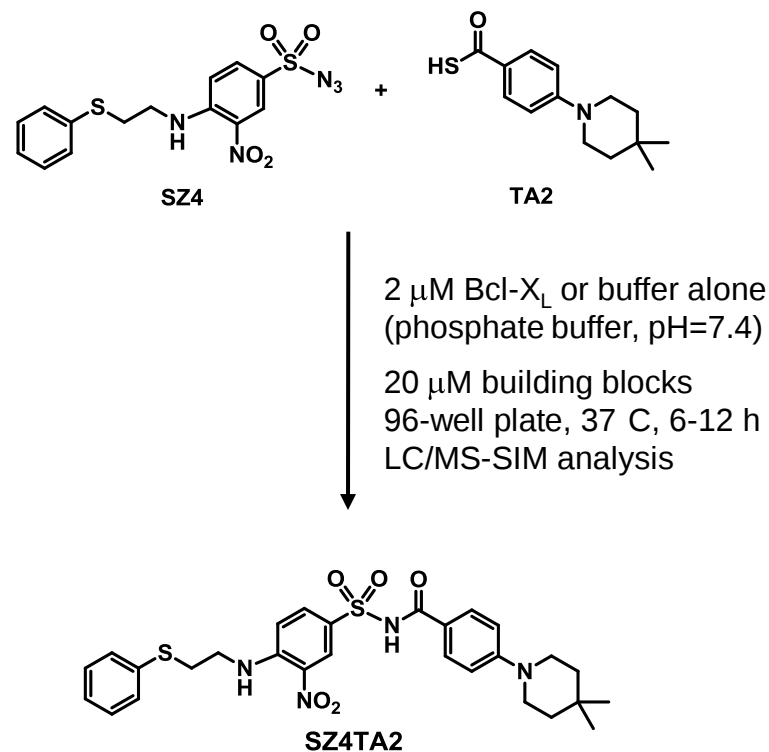
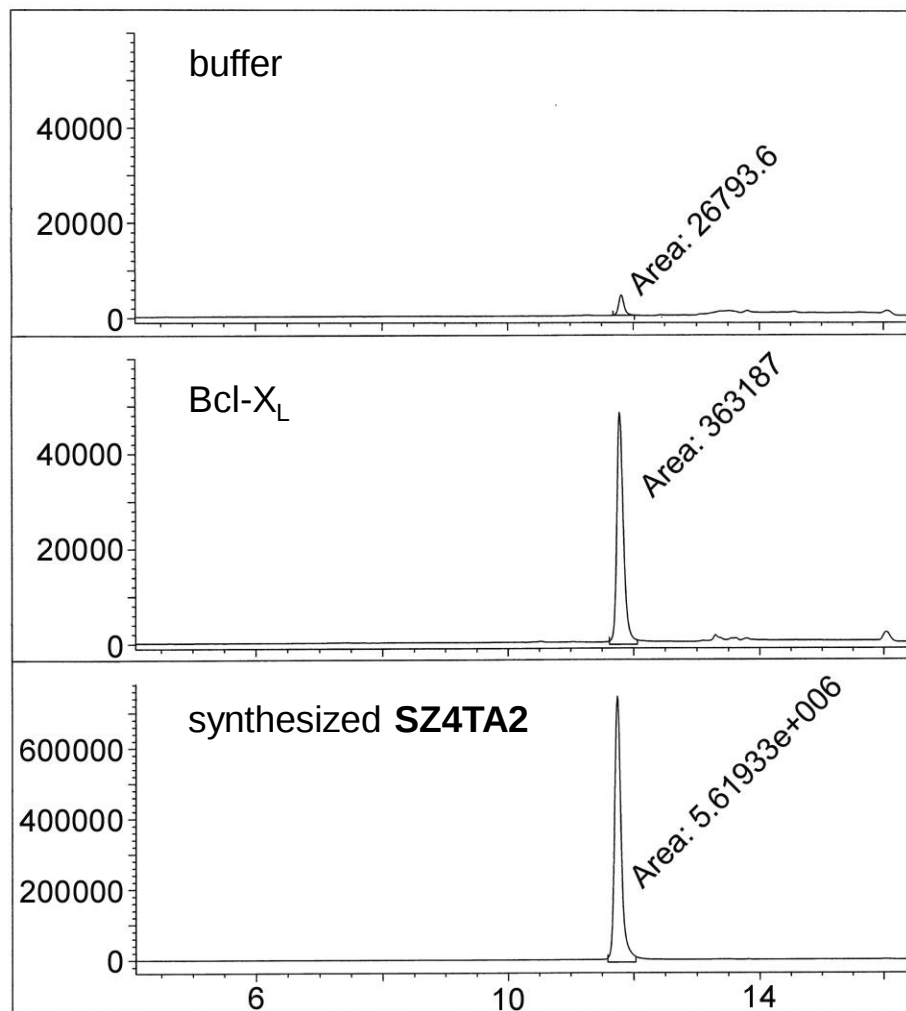
Scan Mode



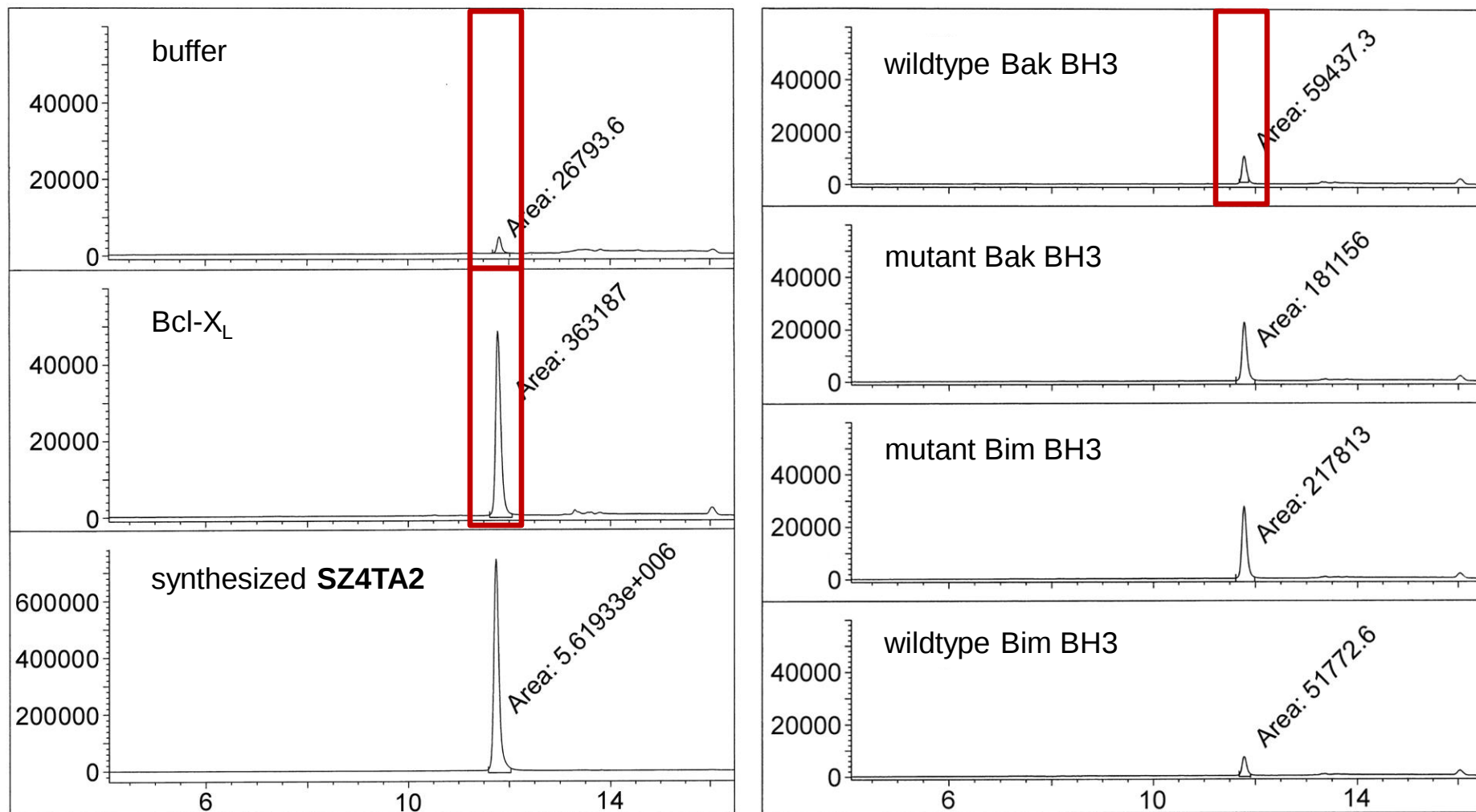
Selected Ion Mode



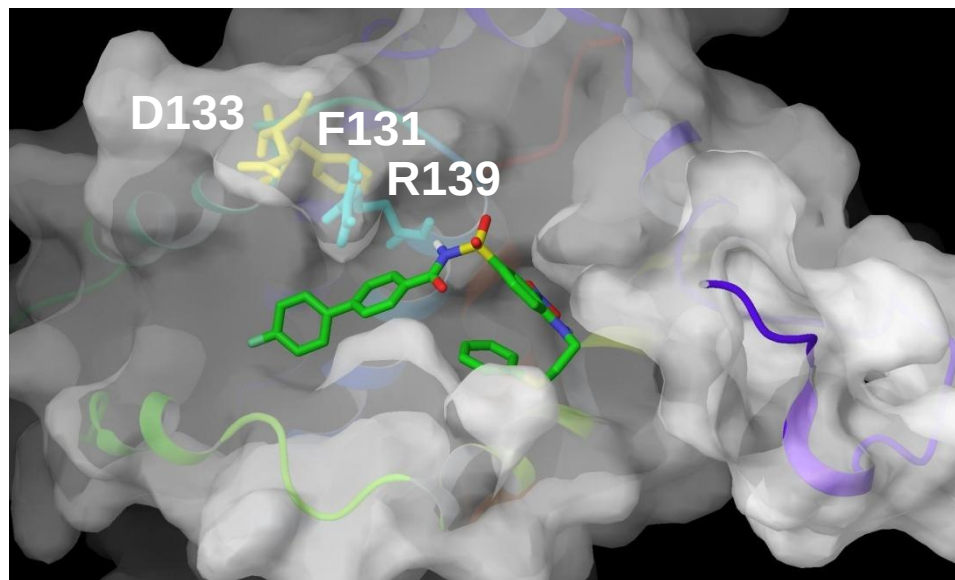
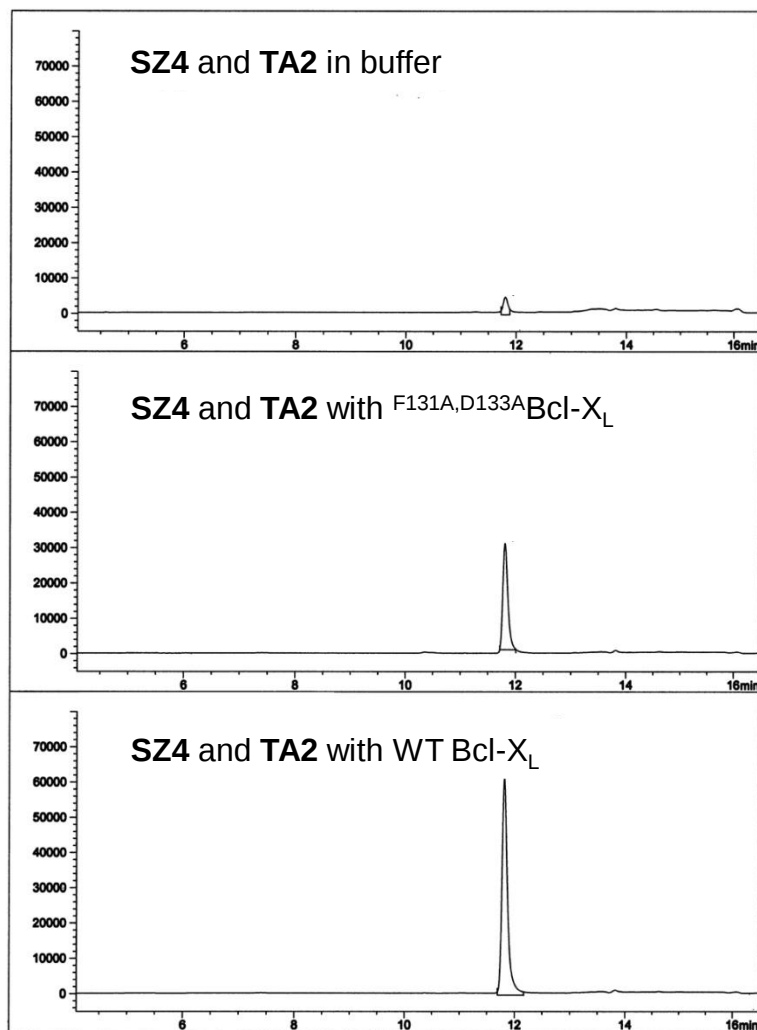
LC/MS-SIM Analysis



Control Incubations: Competition with Bim or Bak



Mutants: Phe131, Asp133 or Arg139 substituted by Ala



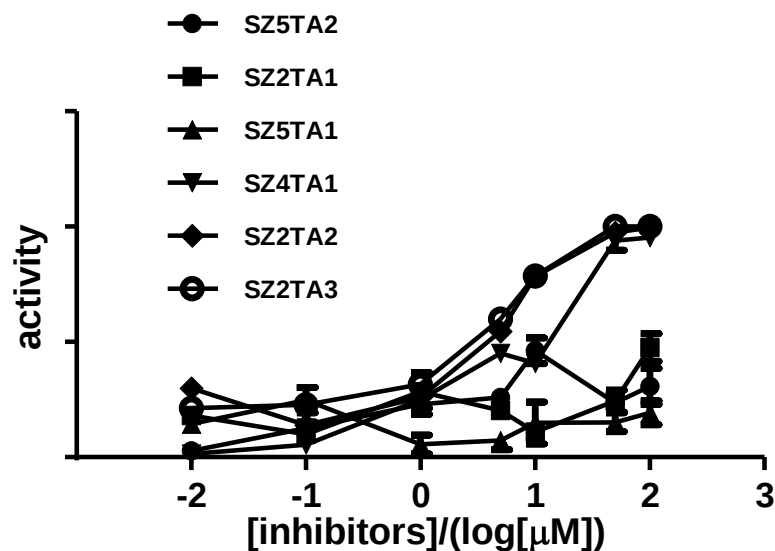
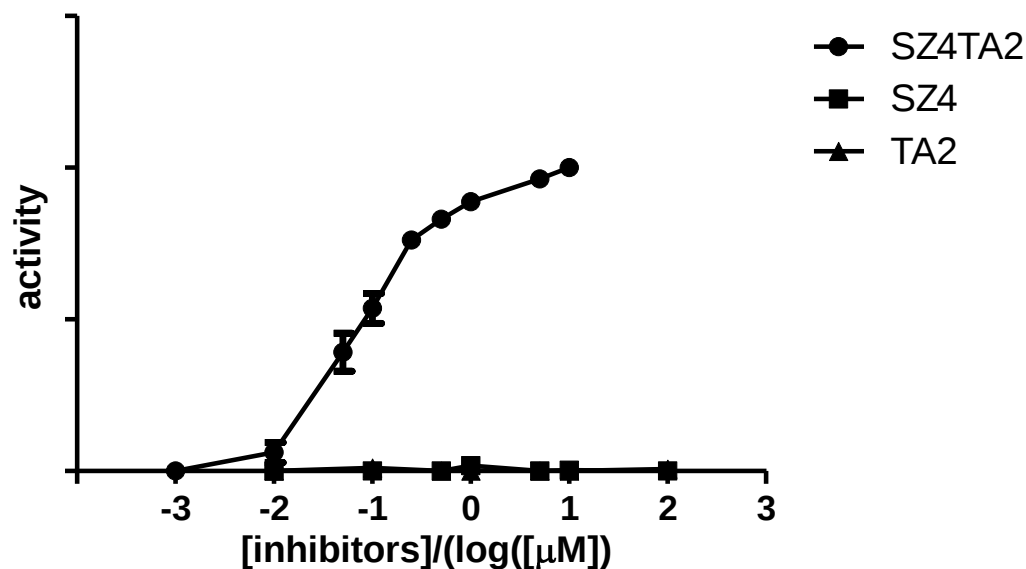
Incubations of **SZ4** and **TA2** with WT or mutant Bcl-X_L and corresponding control incubations with Bak or Bim BH3 peptides.

Entry	Experiment	PA [#]	% Signal
1	Buffer alone	26,794	7.4
2	WT Bcl-X _L	363,187	100.0
3	WT Bcl-X _L and WT Bak	59,437	16.3
4	WT Bcl-X _L and mutant Bak	181,156	49.8
5	WT Bcl-X _L and WT Bim	51,773	14.3
6	WT Bcl-X _L and mutant Bim	217,813	59.9
7	F131A,D133ABcl-X _L	157,059	43.2
8	R139ABcl-X _L	95,154	26.2

[#] PA = Peak Area



Binding: Fluorescence Polarization Assay

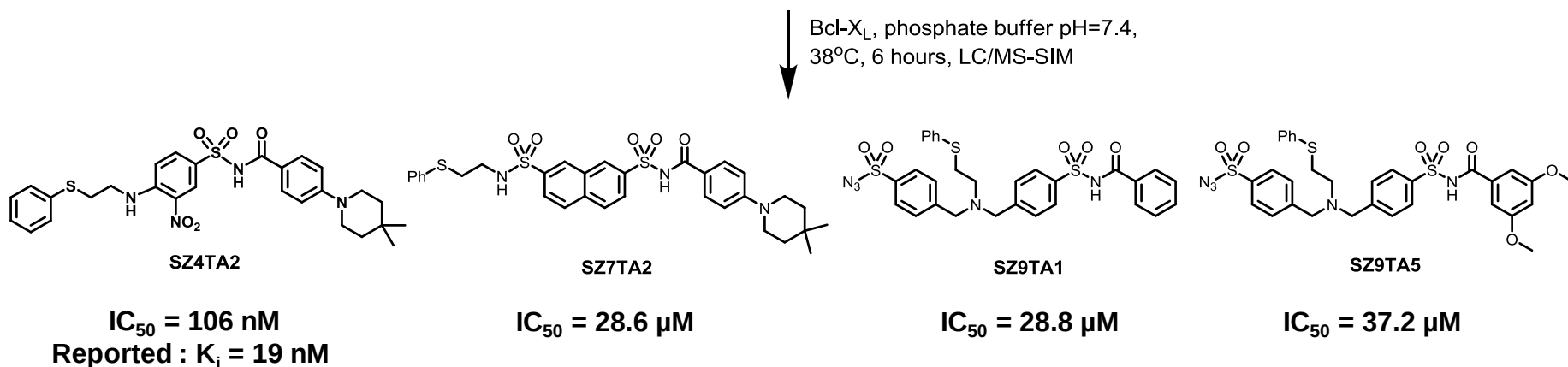
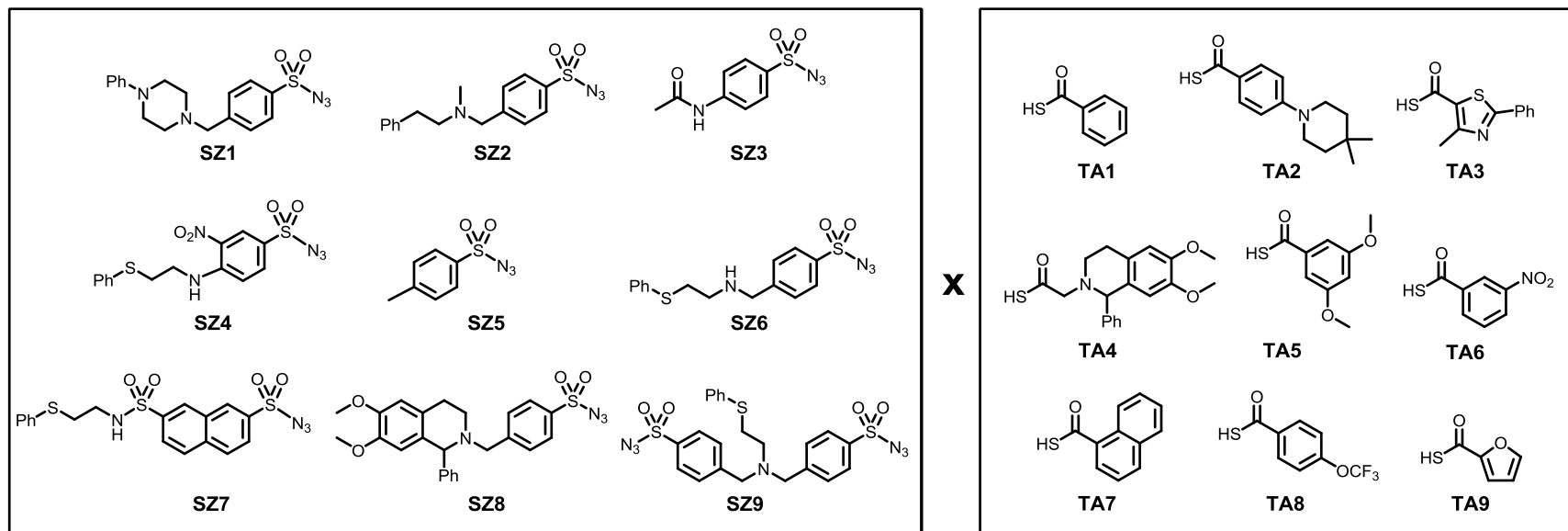


Compound	IC ₅₀ (FP)
SZ4TA2	78.8 6.7 nM
SZ4	>100 μM
TA2	>100 μM
SZ5TA2	>100 μM
SZ2TA1	>100 μM
SZ5TA1	>100 μM
SZ4TA1	38.9 8.4 μM
SZ2TA2	5.19 1.96 μM
SZ2TA3	5.34 4.24 μM

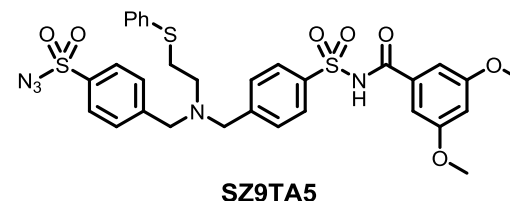
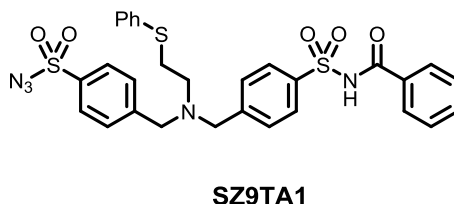
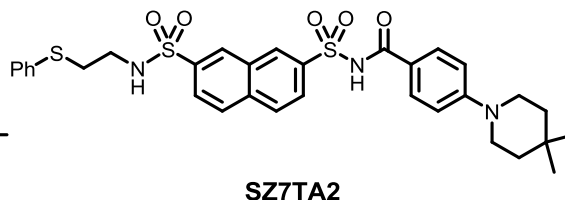
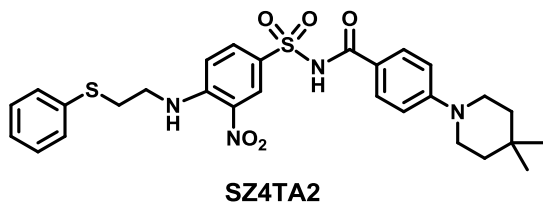
Bcl-X_L-Templated Reactions and Screening



Total 18 reactive building blocks: 9 sulfonyl azides and 9 thio acids leading to 81 potential amides (9 x 9 = 81)



Are TGS Hit Compounds the Most Potent Compounds?

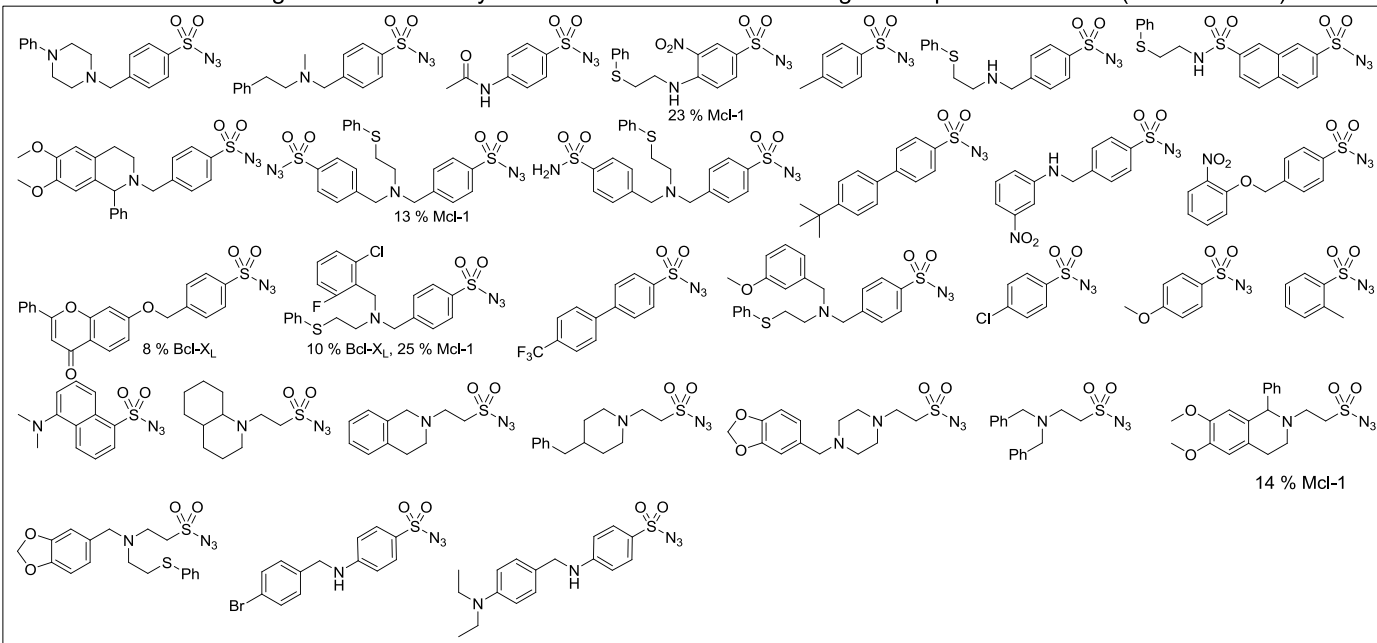


Fragments	SZ1	SZ2	SZ3	SZ4	SZ5	SZ6	SZ7	SZ8	SZ9
TA1	nd	2	0	14	29	nd	nd	19	80
TA2	nd	9	nd	100	28	26	76	nd	38
TA3	6	7	nd	nd	nd	nd	nd	30	22
TA4	nd	25	nd	nd	nd	nd	nd	8	nd
TA5	5	nd	nd	nd	0	nd	15	11	60
TA6	4	nd	0	nd	0	nd	20	nd	46
TA7	nd	nd	0	nd	nd	nd	47	30	nd
TA8	3	nd	0	nd	nd	nd	nd	38	nd
TA9	nd	nd	0	nd	1	nd	nd	24	nd

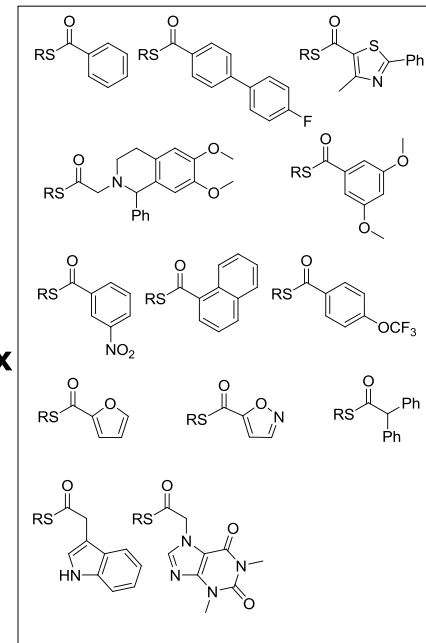
% inhibition at 50 μ M acylsulfonamide concentration; 37 compounds synthesized and tested

Parallel Kinetic TGS Screening of Multiple PPIs

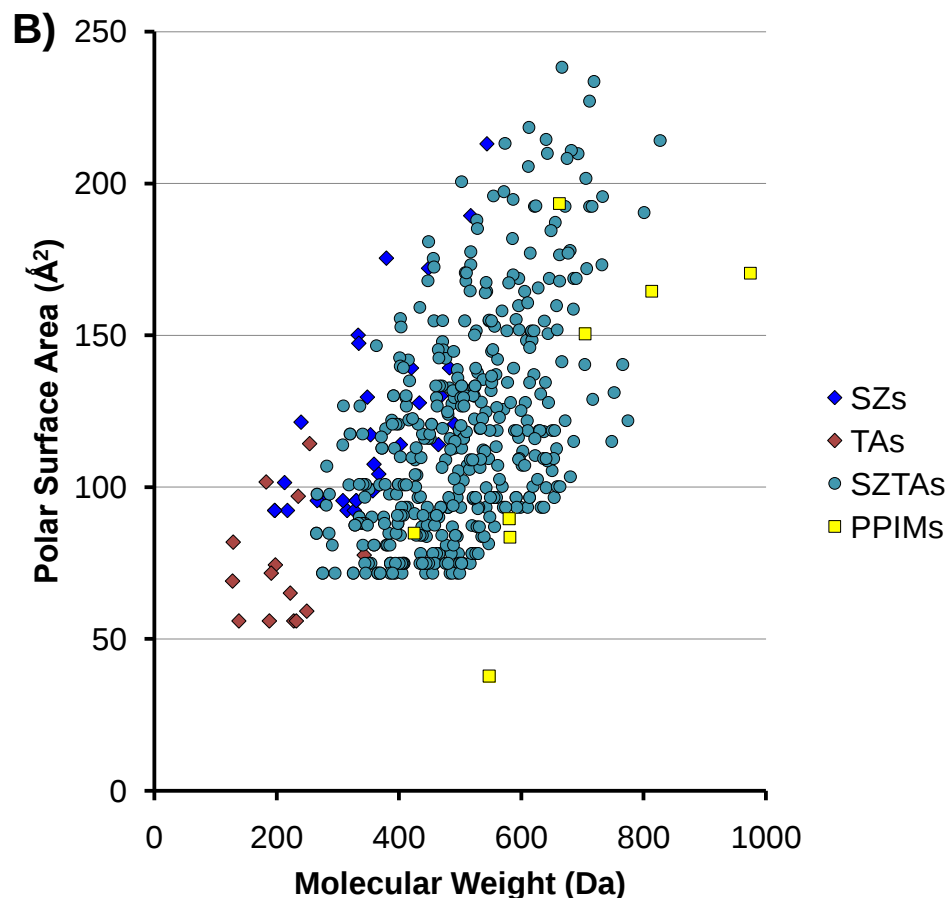
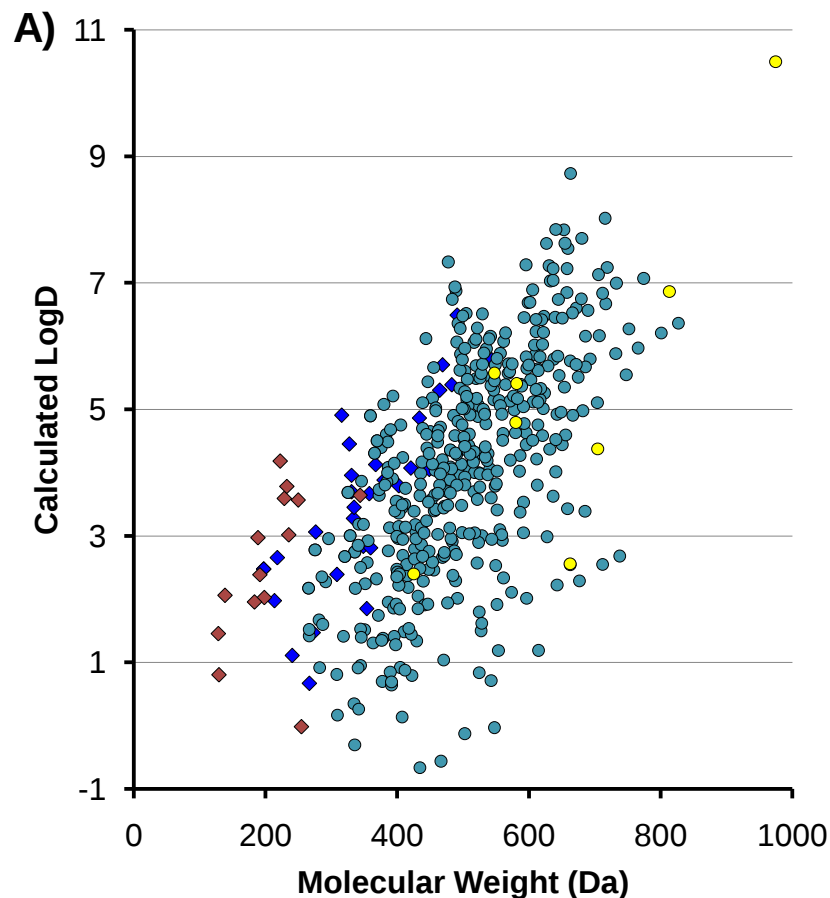
Total 43 reactive building blocks: 30 sulfonyl azides and 13 thio acids leading to 390 potential amides (30 x 13 = 390)



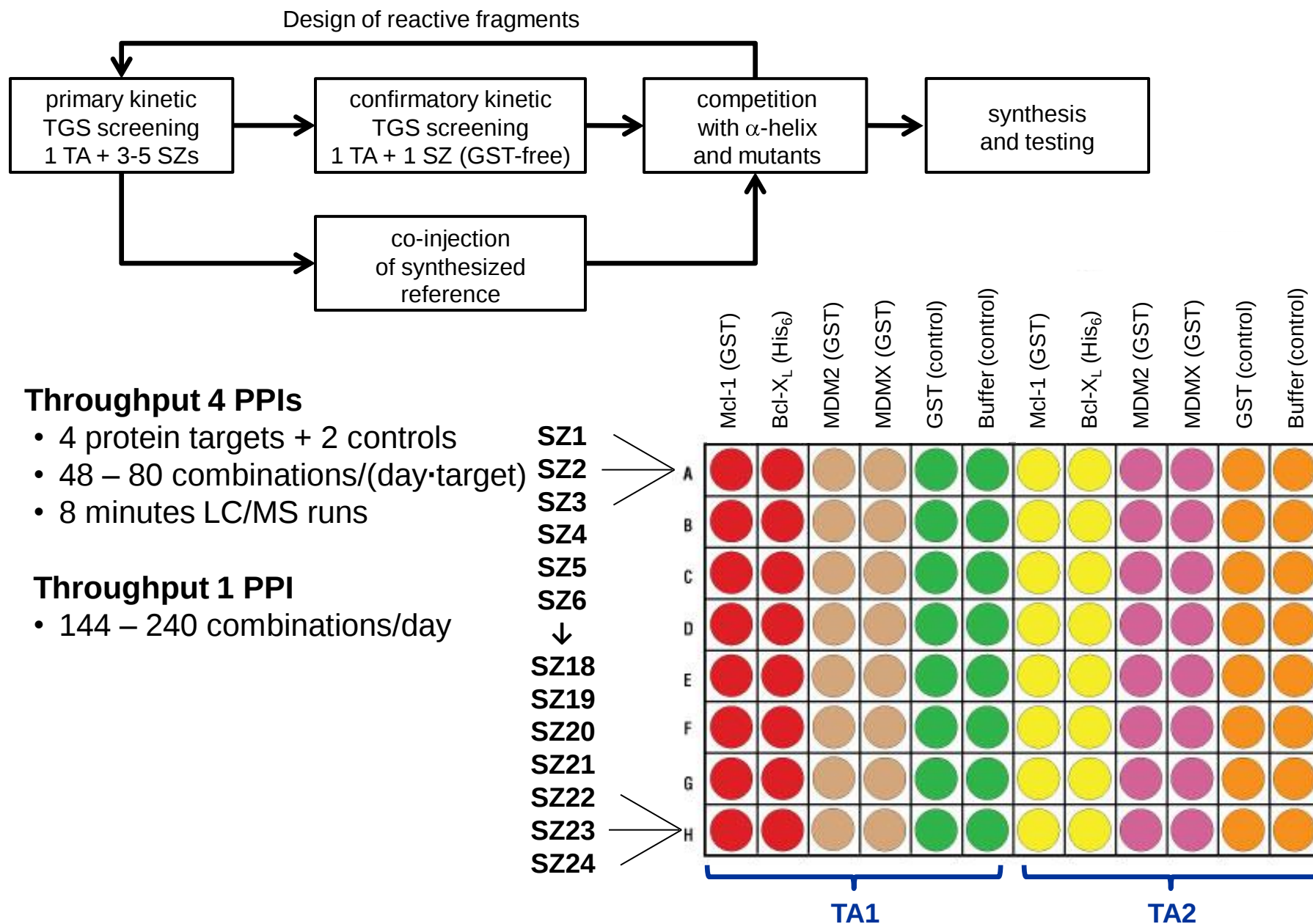
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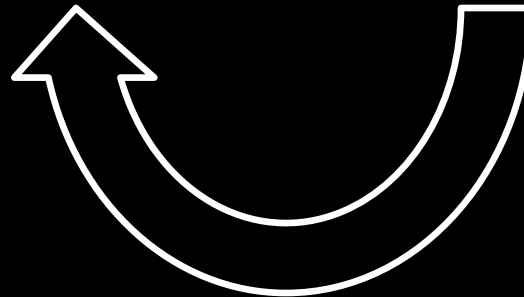
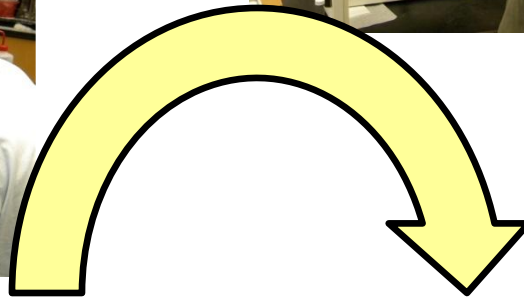
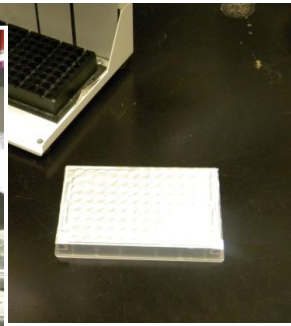
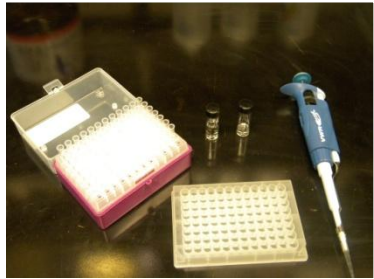
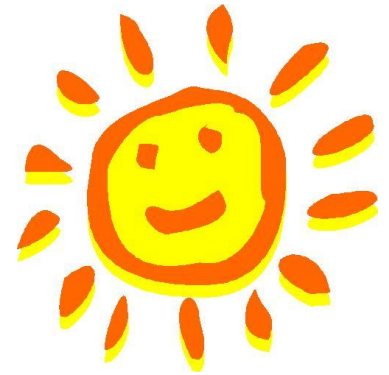
Building Blocks and Acylsulfonamides



Parallel Kinetic TGS Screening of Multiple PPIs

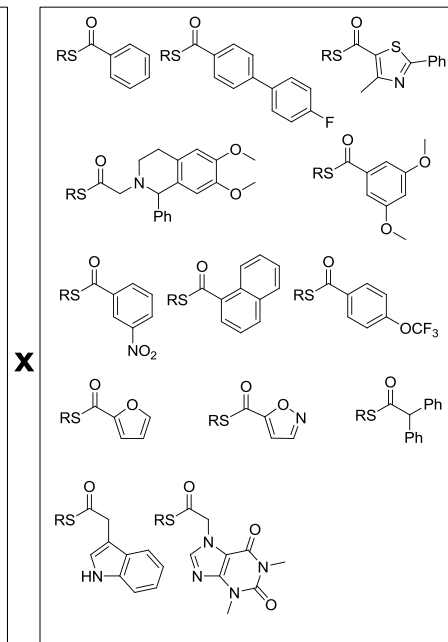
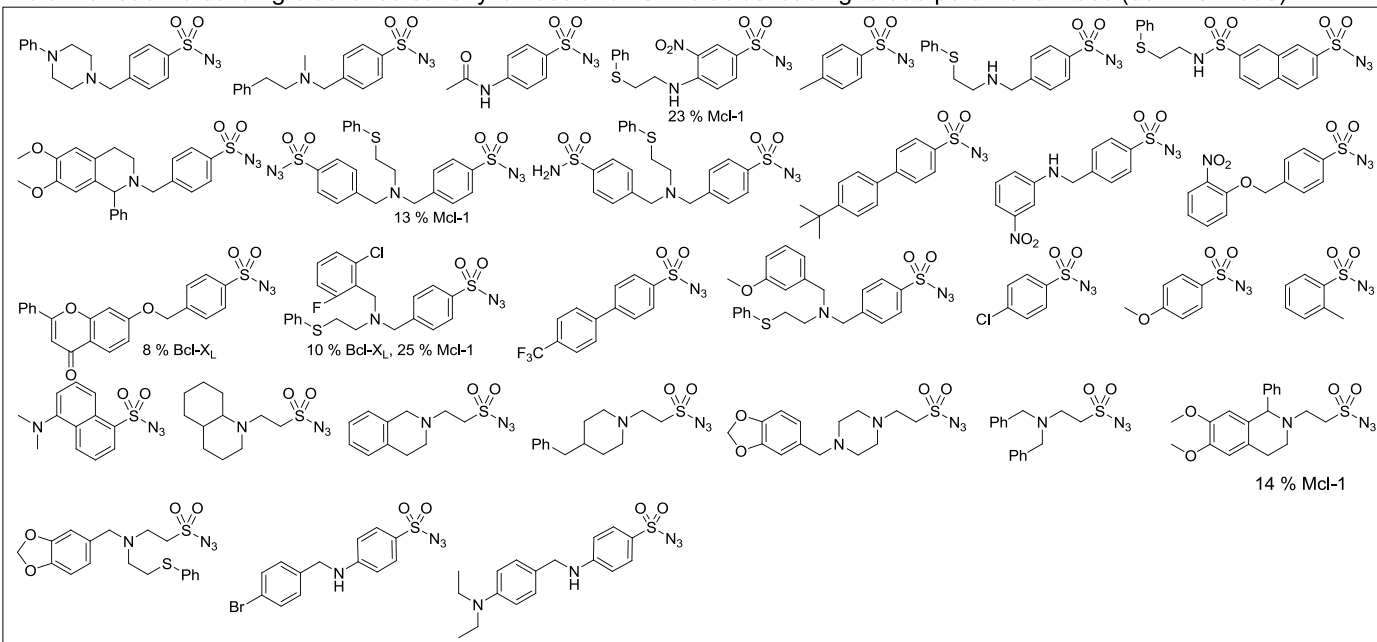


Parallel Kinetic TGS Screening of Multiple PPIs



Parallel Kinetic TGS Screening: Example Mcl-1

Total 43 reactive building blocks: 30 sulfonyl azides and 13 thio acids leading to 390 potential amides (30 x 13 = 390)



Incubation conditions: 8 μ M protein (phosphate buffer, pH=7.4)
 20 μ M sulfonylazide and 20 μ M thioacid building blocks
 Mixtures of 1 thio acid and 3-5 azides, 37 C, 12 h
 LC/MS-SIM analysis

Example: $\pm$ 500 combinations $\rightarrow$ 13 kinetic TGS hits for Mcl-1 $\rightarrow$ 1 SZ class



Conclusions

- Amidation between thioacids and sulfonylazides can be utilized for kinetic TGS
- Control incubations to confirm that kinetic TGS occurs at the hot spots of Bcl-X_L, Mcl-1, or any other PPIs
- Kinetic TGS hit acylsulfonamides are more potent compared to acylsulfonamides not discovered through kinetic TGS
- Straightforward and rapid “in-well” activation of thioesters to yield thioacids
- Kinetic TGS screening lead to the discovery of selective acylsulfonylamide inhibitors targeting Mcl-1 over Bcl-X_L



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Kinetic TGS

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