

Biophysical ligand screening for soluble and membrane proteins

September 2010
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Leiden, The Netherlands

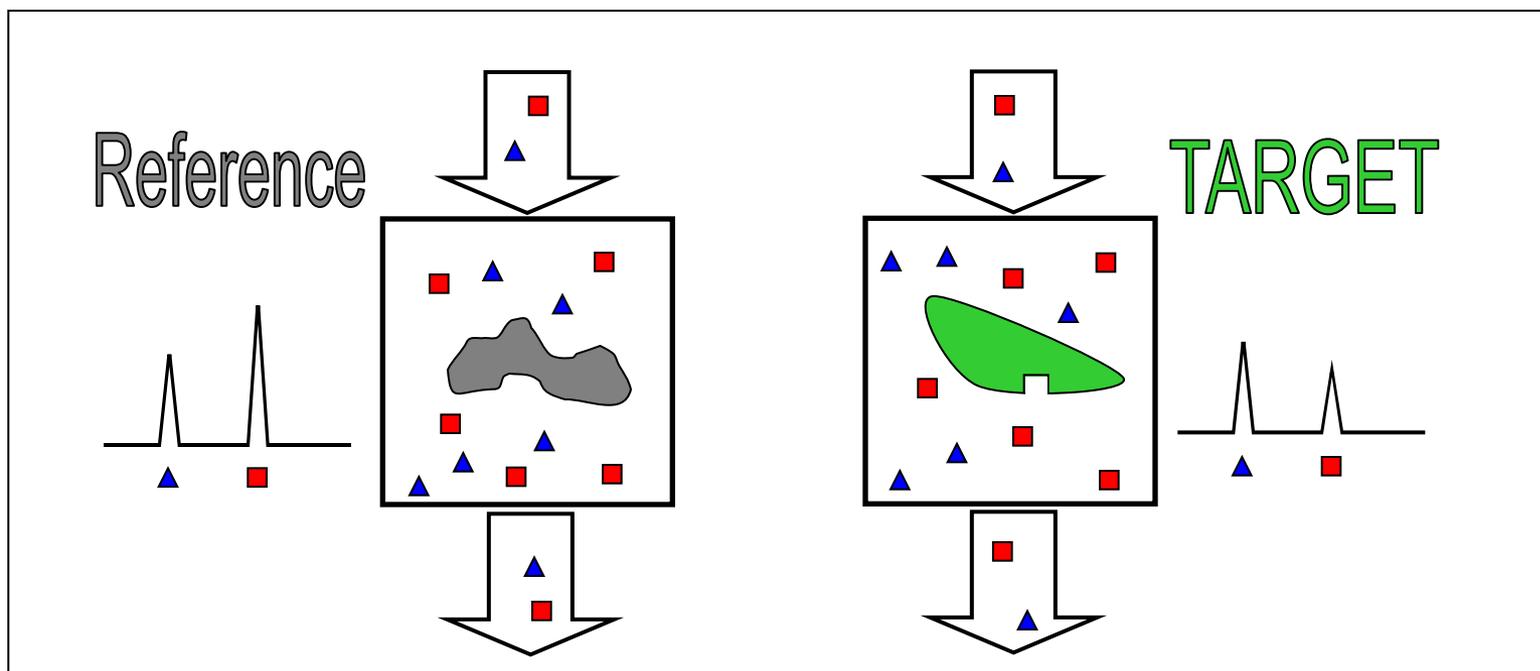


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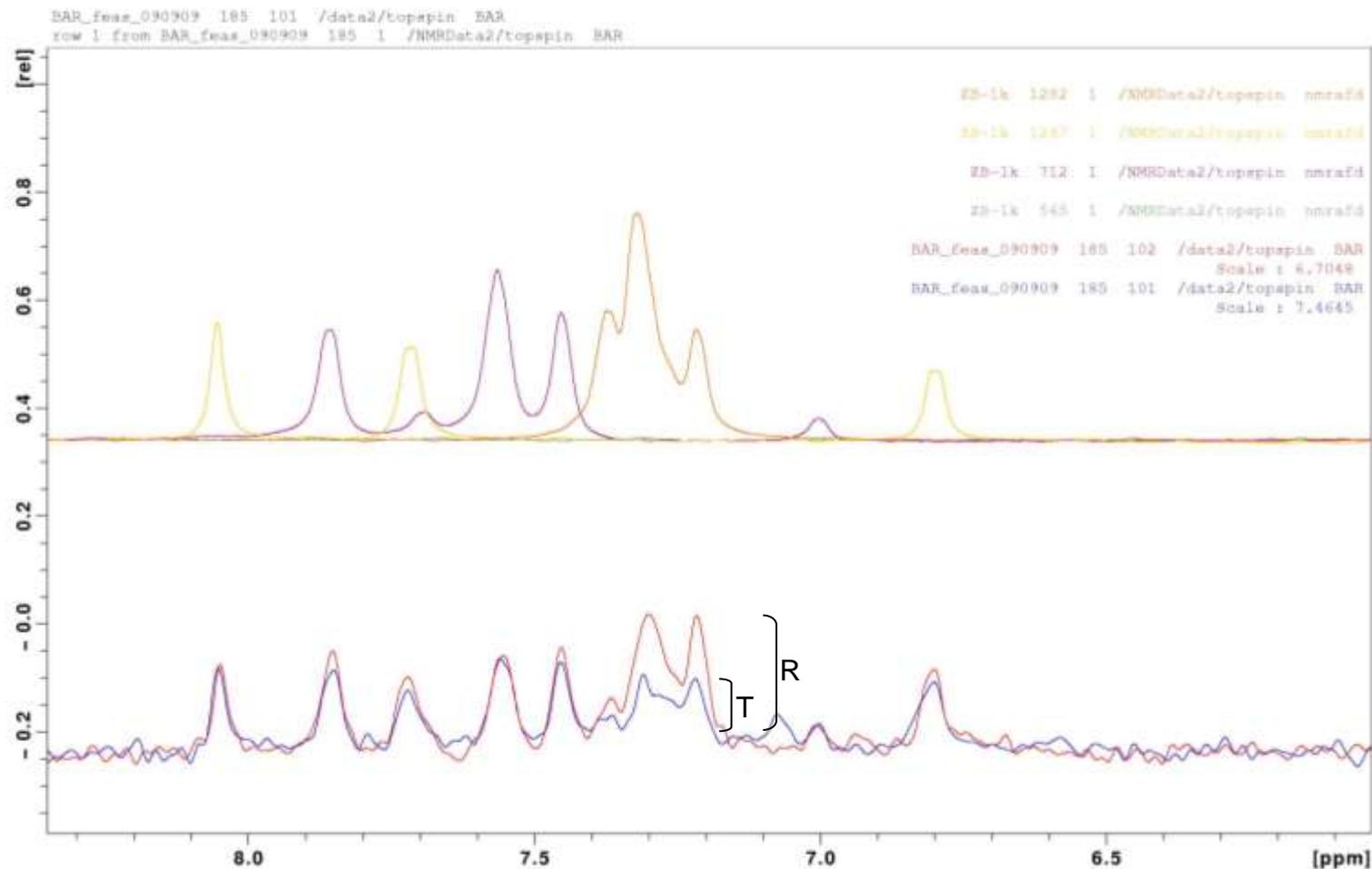
The TINS method for finding hits

TINS = Target Immobilized NMR Screening



Vanwetswinkel et al., 2005,12, p.207

An example of raw data



Summary of Selected Immobilized Targets

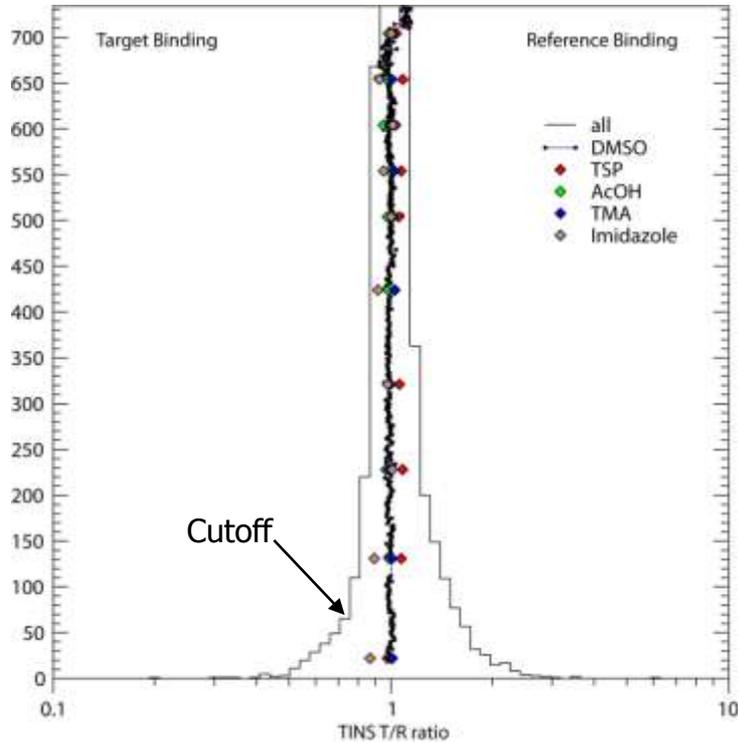
Protein	Size	Functional	TINS hits
Protease	44 kDa	LB	5.9%
HSP90	24 kDa	LB	6.5%
Small GTPase	20 kDa	LB/BA	9% apo form 3% NDP form
Viral enzyme	67 kDa	LB/BA	9.5%
DsbB (Bacterial mem. Prot.)	14 kDa	BA	7.3%
Various kinases (pY,pS/T)	30-35 kDa	LB/BA	3.8-5.1%
KcsA (Ion channel)	57 kDa	LB	Feasibility only (95 cmpds, 7%)
Metalloproteins	105 kDa homotrimer	LB/BA	5-8.5%
Prot-Prot Interaction (5)	14-100 kDa	LB	3-6%

LB ligand binding
BA biochemical assay

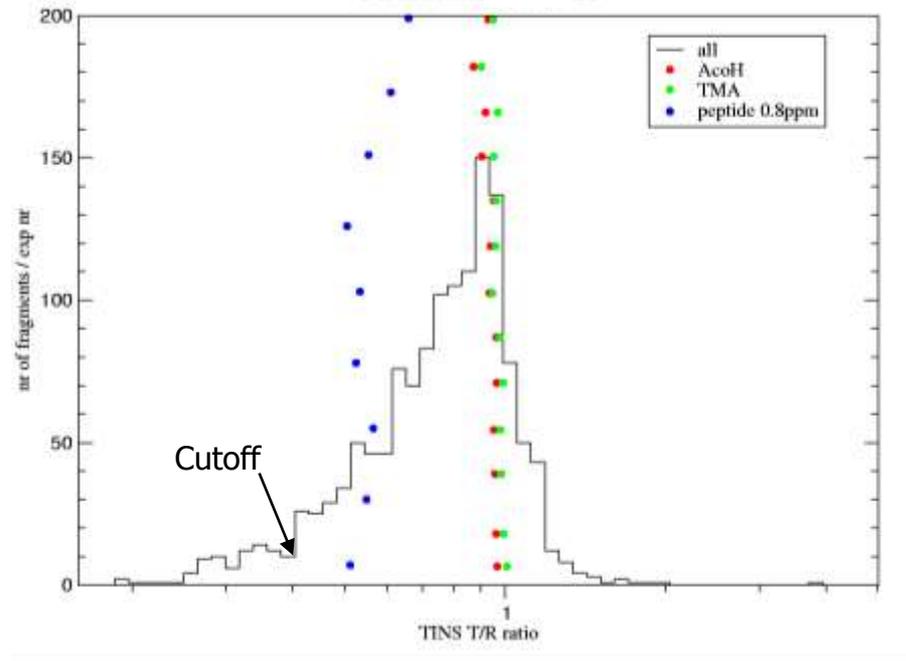
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Fragment screening with TINS



Undruggable Target
11 hits

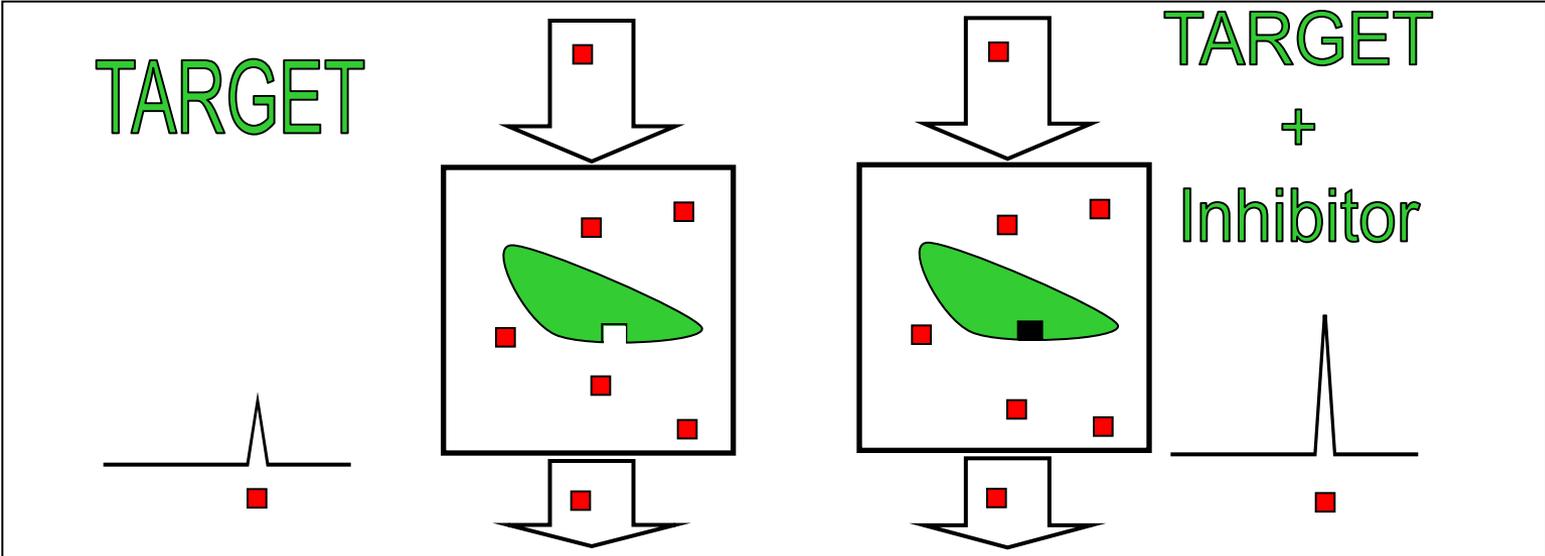


Druggable Target
89 hits

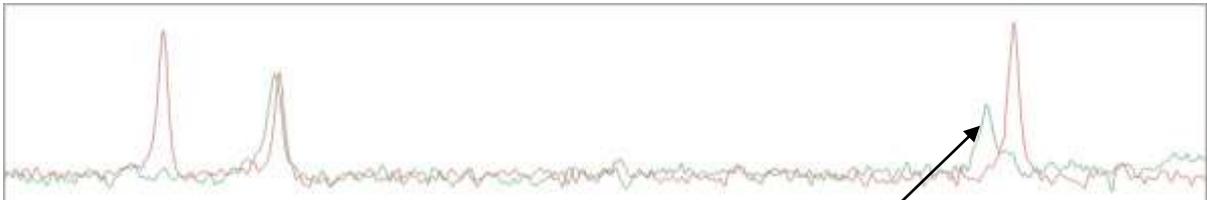
Methods for Hit Validation

- Affinity ranking
- Competition Binding
- SPR
- HSQC Binding Site Determination

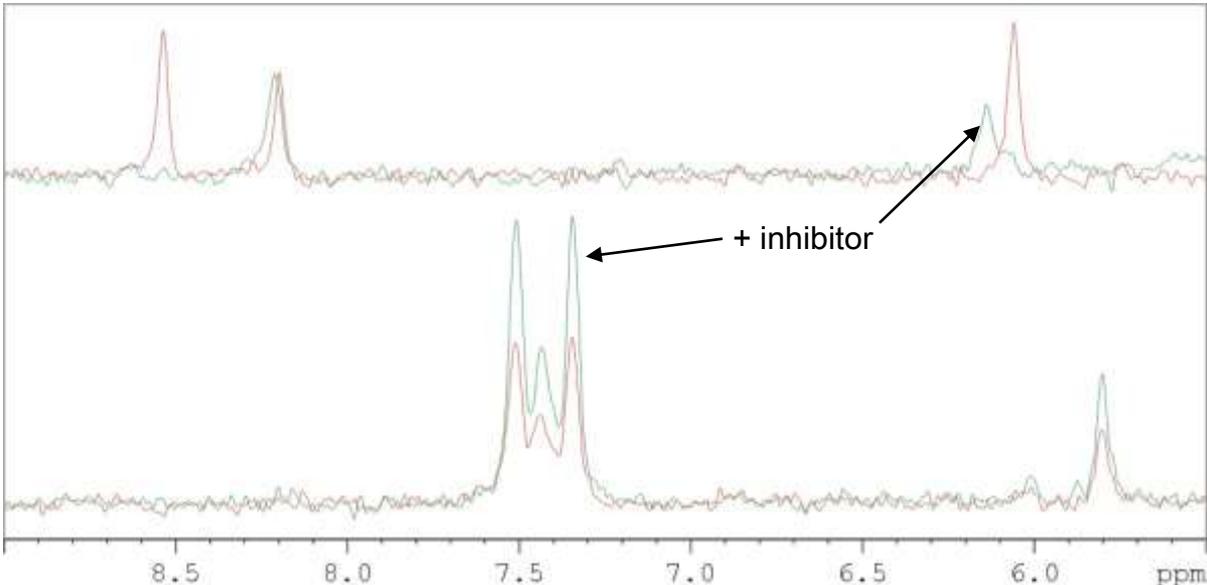
Hit Validation: Competition Binding in TINS



Biological Assay

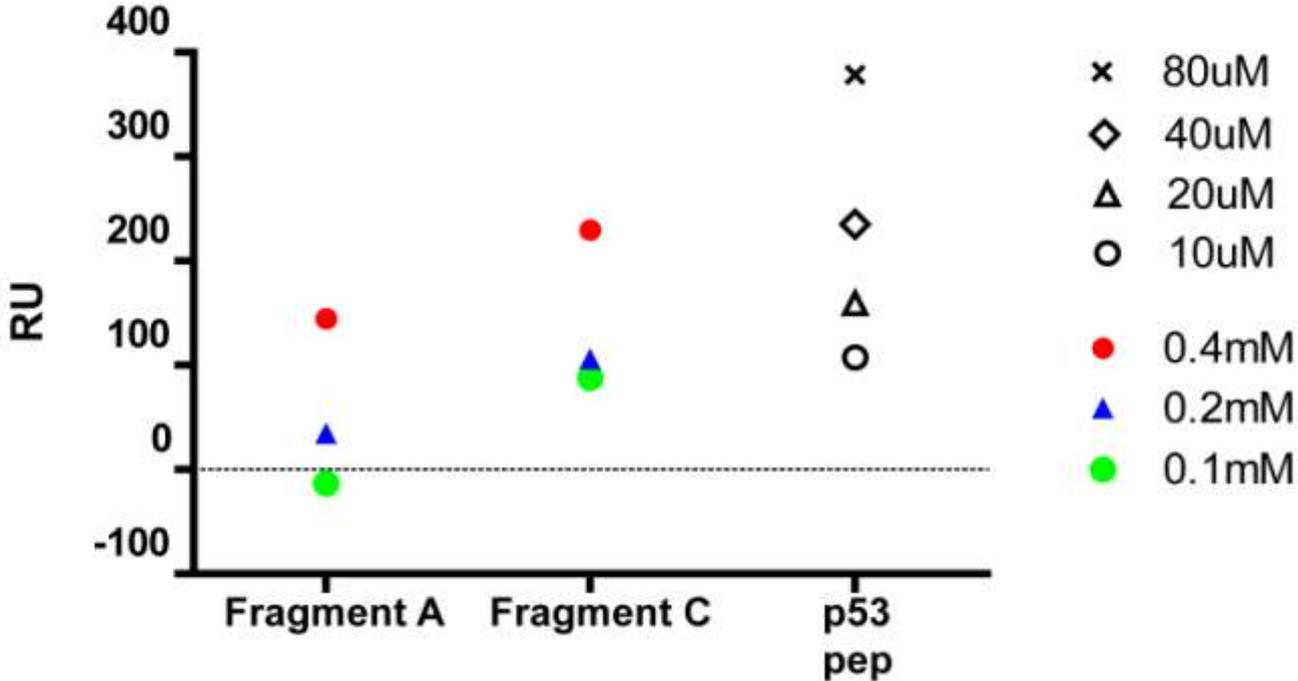


Biophysical Assay



Hit Validation: SPR (Biacore T200)

Biacore Analysis of two small molecules binding to MDM4

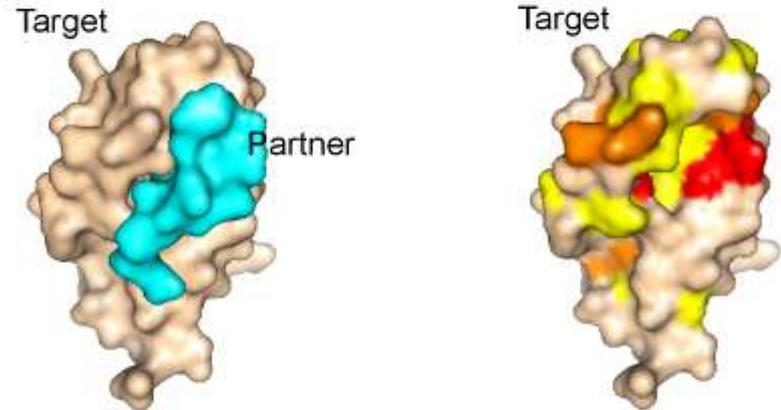


Hit Validation: K_D and Binding Site Characterization by HSQC

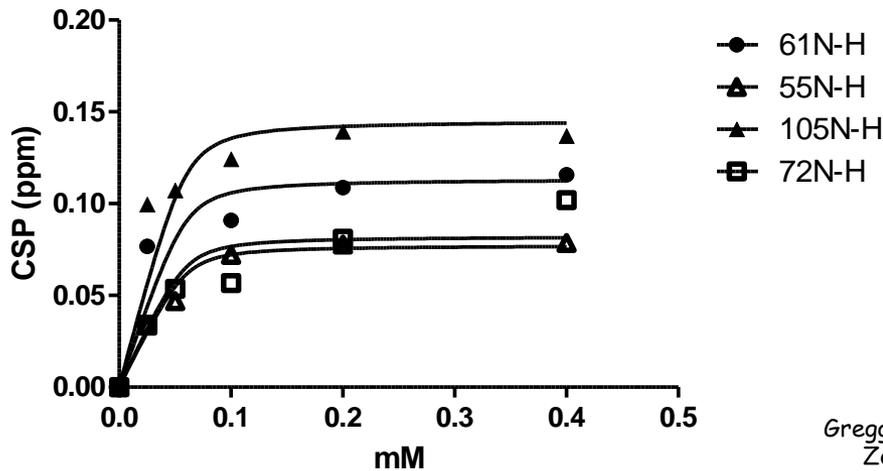
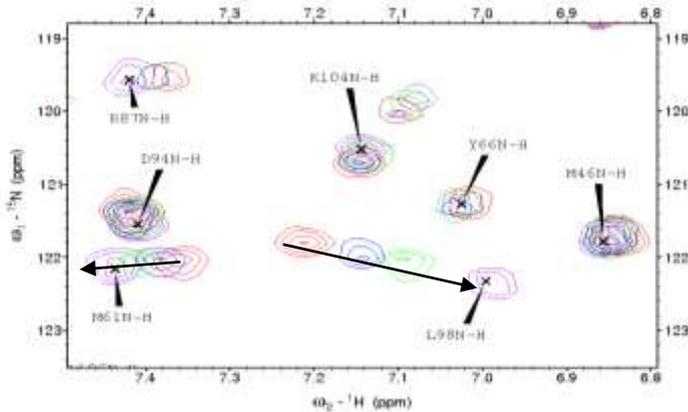
- Chemical shift perturbation for each HN
 - magnitude of resonance shift due to compound binding
- $CSP = \sqrt{[\Delta H_{ppm}]^2 + (\Delta N_{ppm}/6.5)^2}$

CSP observation on 80 TINS Hits

Low resolution binding site determination



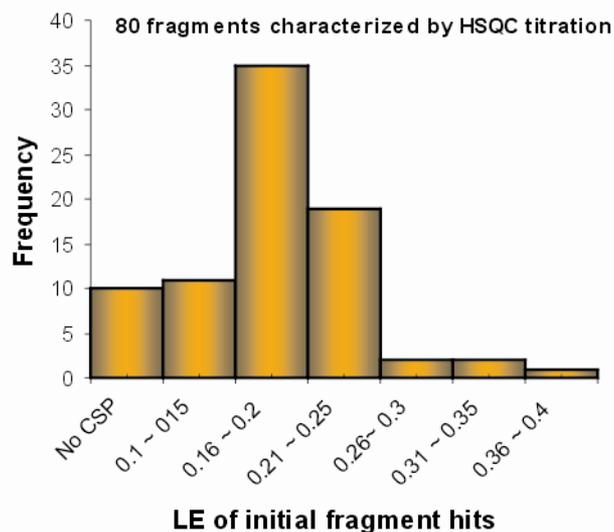
Hits bind in the vicinity of PPI site



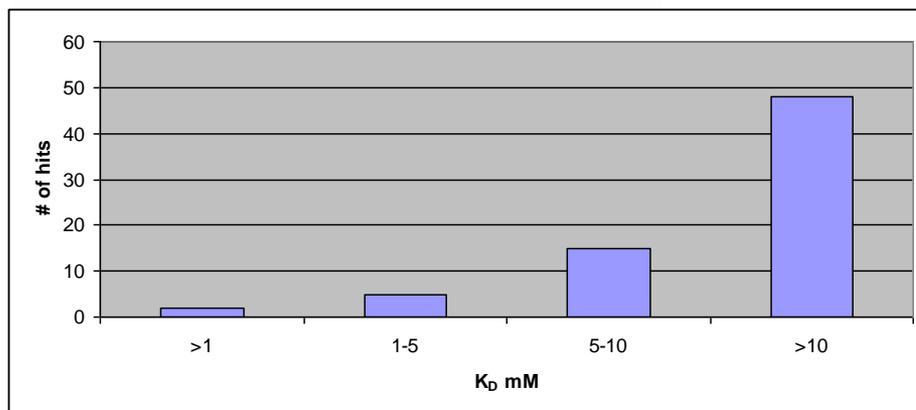
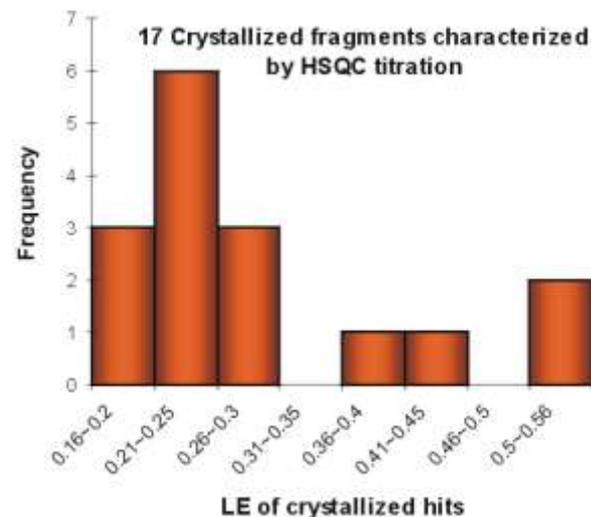
Characteristics of Fragment Hits from TINS

All TINS hits were assessed by [^{15}N , ^1H]-HSQC titration

Against PPI



Against ATPase (HSP90)*

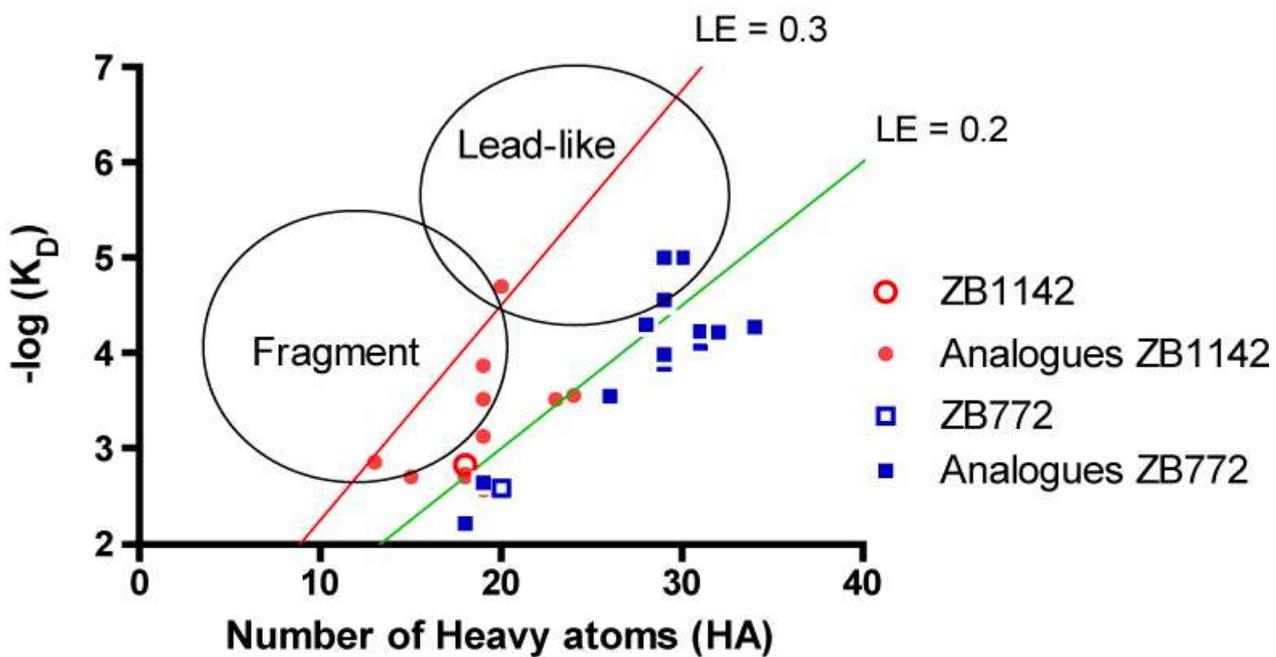


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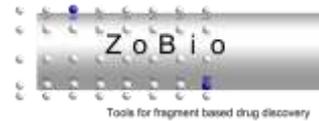
Elaboration of TINS hits guided by K_D from NMR titration

- X- ray crystallography failed to obtain protein – fragment complex
- Binding site and K_D determined by HSQC titration supported ligand-target docking

Analoging of TINS hit against PPI



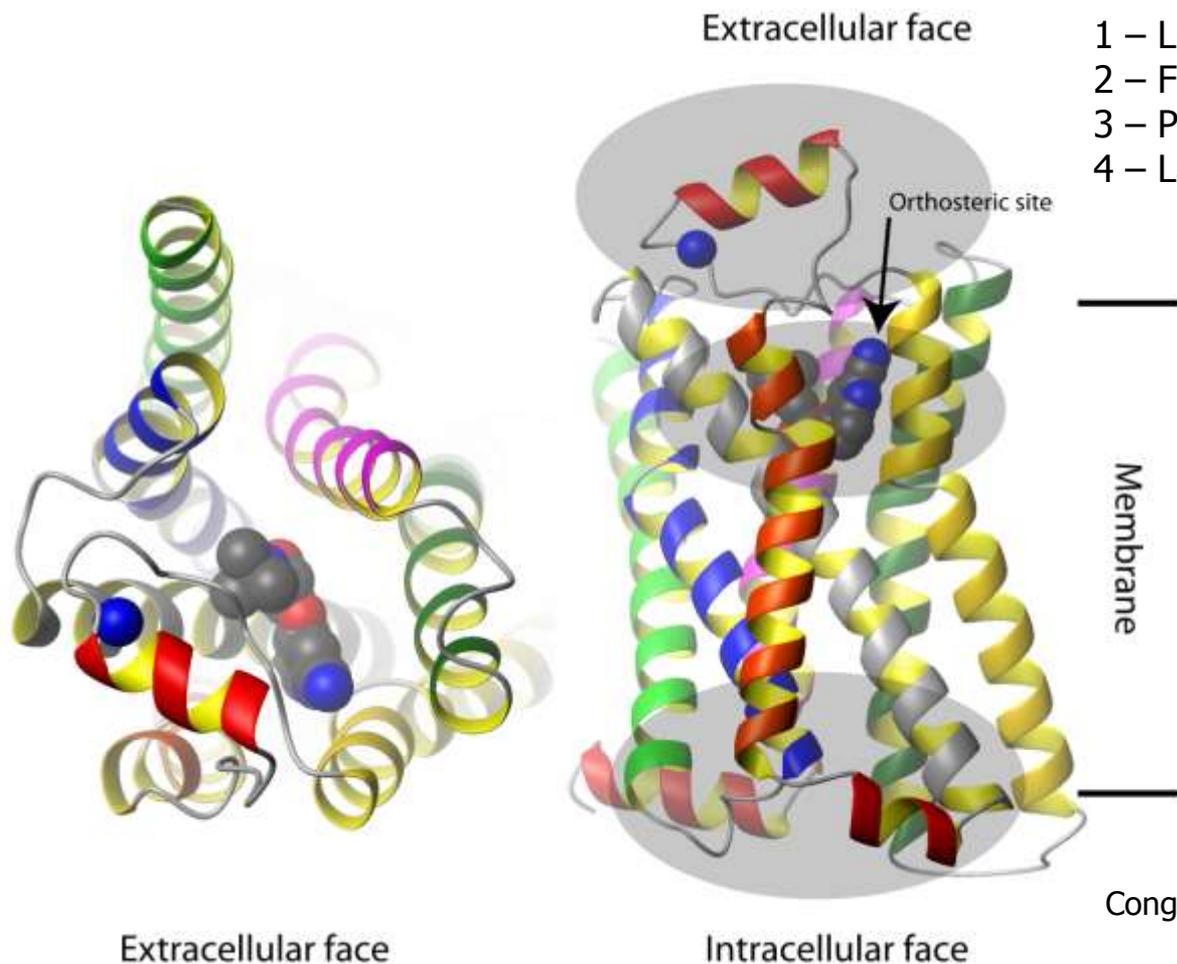
Good starting point as a Lead-like molecule



The Challenges of Fragment Discovery on MPs

- Protein production/solubilization/stability
- Non-specific binding
- Slow kinetics
- Fragment size

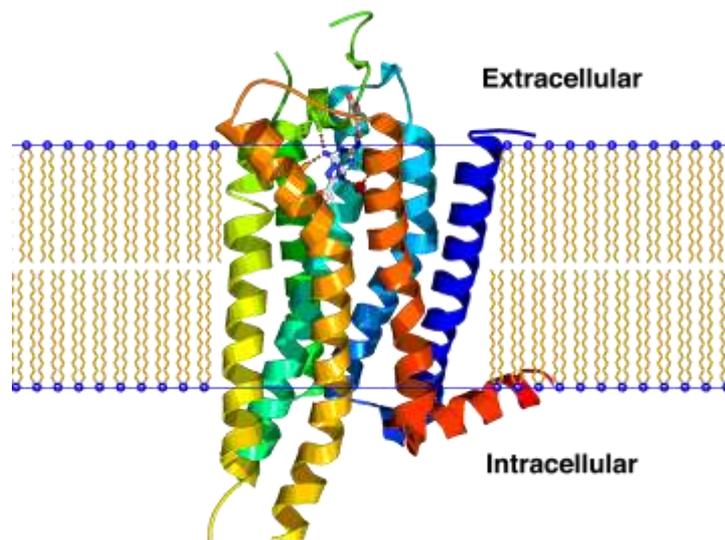
Why biophysical fragment screening for GPCRs?



- 1 – Ligands for peptide activated GPCRs
- 2 – Find sites with novel biological function
- 3 – Provide structural information
- 4 – Label free

Congreve et al., Methods in Enzymology, in press

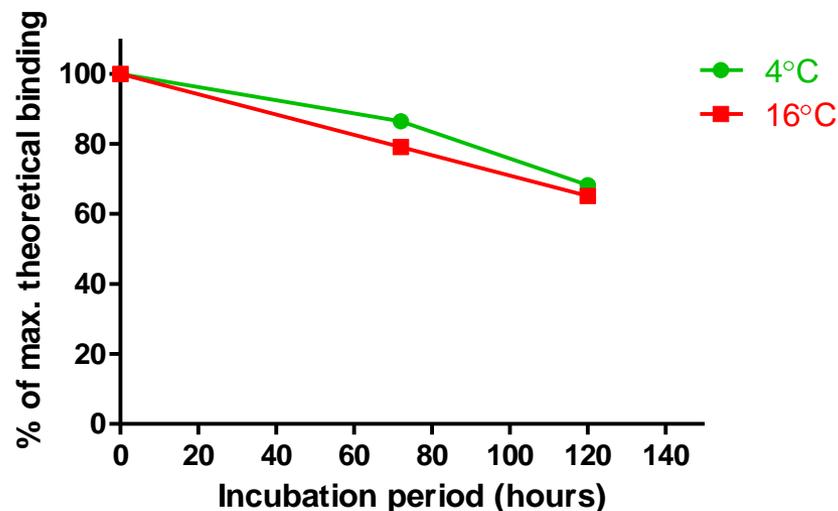
Fragment Screening of GPCRs: Adenosine 2a & β 1 Adrenergic receptors



$A_{2a}R$ agonists – anti-inflammatory therapeutic potential

$A_{2a}R$ antagonists – used to treat Parkinson's disease as $A_{2a}R$ dimerises with dopamine D_2 receptor

Radioligand binding on immobilized adenosine A_{2a} receptor



Immobilized hA2aR maintained 60% the activity in five days.

Profile of Ligand Binding in the Screen

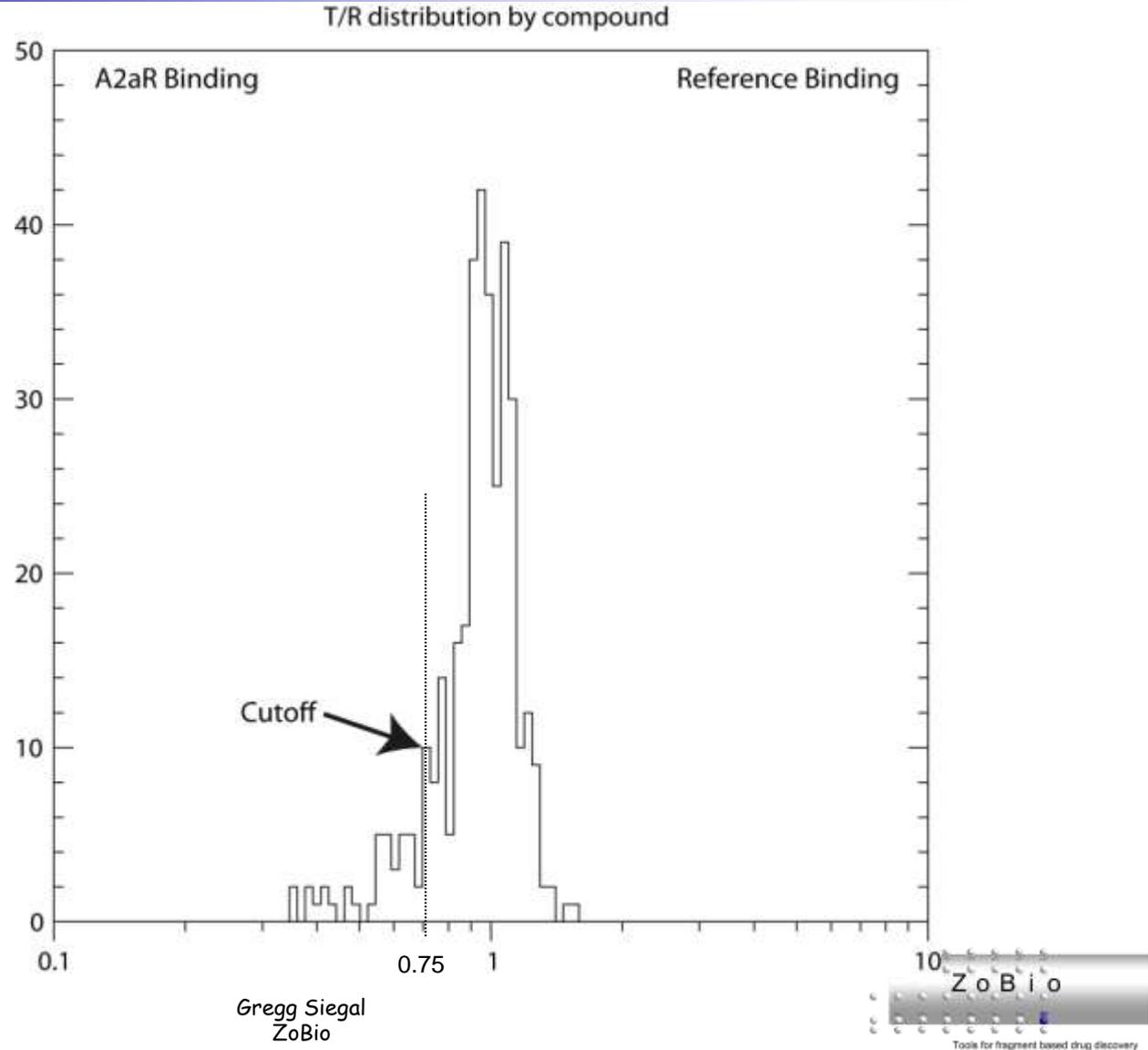
531 fragments assayed
against **apo** A2aR

94 hits with very loose
selection parameters

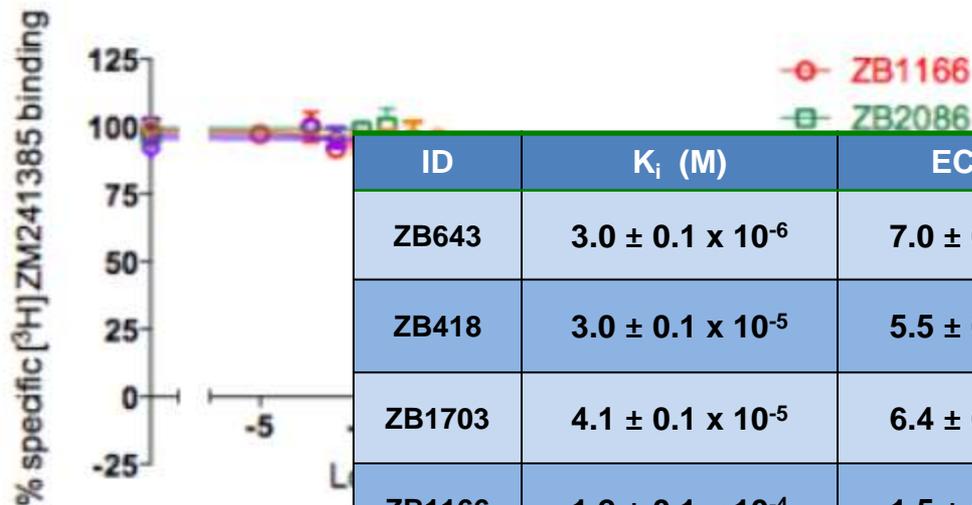
β 1AR:
579 fragments screened
73 hits with a cut-off at T/R = 0.65
Congreve *et al*, MiE, in press

A2aR:
531 fragments screened
94 hits with a cut-off at T/R = 0.75

Number in common: 20 hits



Hit validation by equilibrium radioligand displacement

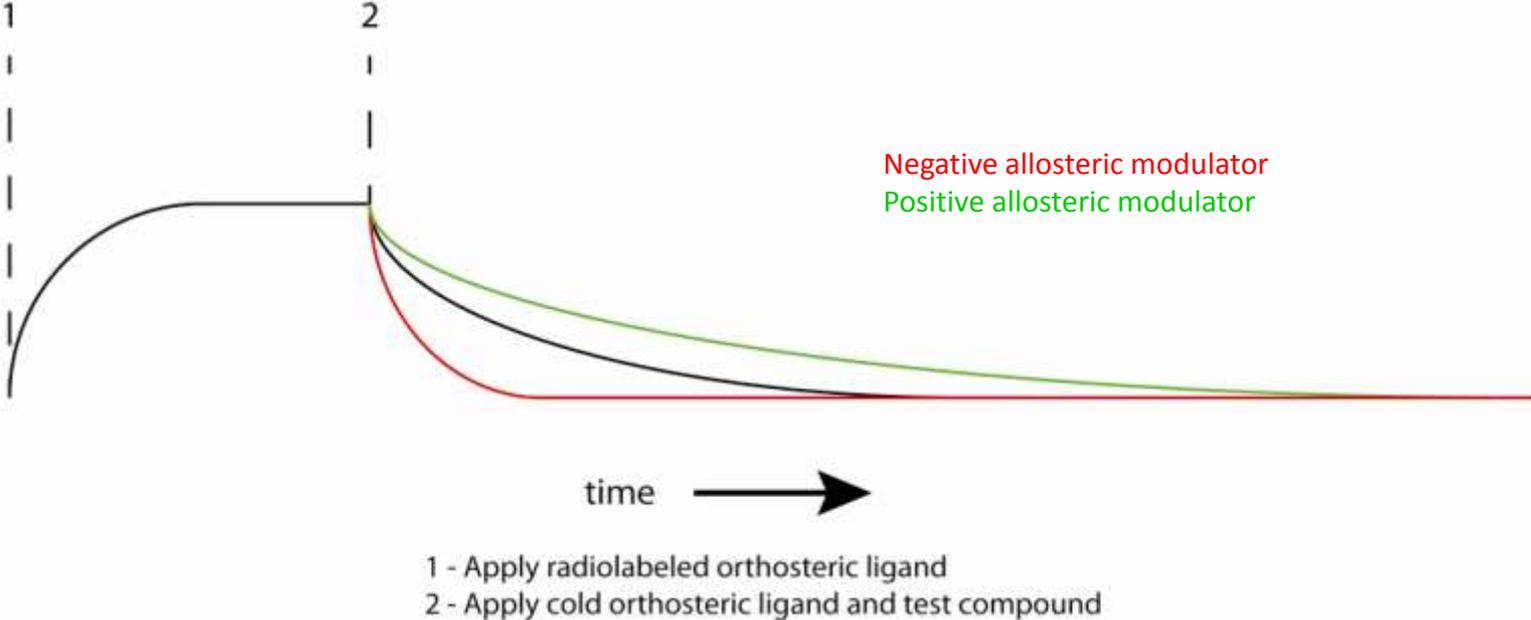


ID	K_i (M)	EC_{50} (M)	Hill Slope	T/R ratio	BEI
ZB643	$3.0 \pm 0.1 \times 10^{-6}$	$7.0 \pm 0.3 \times 10^{-6}$	-0.5 ± 0.1	0.65	26.3
ZB418	$3.0 \pm 0.1 \times 10^{-5}$	$5.5 \pm 0.1 \times 10^{-4}$	-2.0 ± 0.5	0.44	17.6
ZB1703	$4.1 \pm 0.1 \times 10^{-5}$	$6.4 \pm 0.1 \times 10^{-4}$	-1.1 ± 0.4	0.60	12.5
ZB1166	$1.2 \pm 0.1 \times 10^{-4}$	$1.5 \pm 0.1 \times 10^{-3}$	-0.9 ± 0.1	0.61	11.6
ZB2086	$1.2 \pm 0.2 \times 10^{-4}$	$2.4 \pm 0.3 \times 10^{-3}$	-0.9 ± 0.3	0.74	16.3
ZB114	$8.2 \pm 0.3 \times 10^{-5}$	$1.2 \pm 0.1 \times 10^{-3}$	-2.2 ± 5.4	0.59	13.5
ZB1605	$8.7 \pm 0.3 \times 10^{-5}$	$1.4 \pm 0.3 \times 10^{-3}$	-1.1 ± 0.5	0.37	12.8
ZB1967	$3.2 \pm 0.2 \times 10^{-4}$	$6.1 \pm 0.1 \times 10^{-3}$	-0.8 ± 1.3	0.61	10.8

Each hit assayed at

385

Mode of action: Orthosteric vs Allosteric modulators



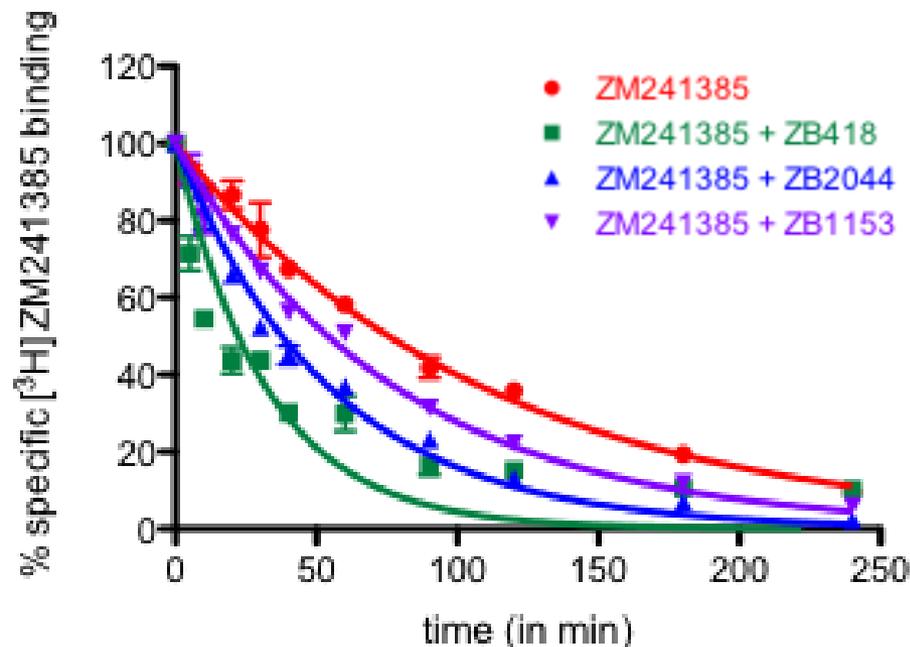
Screen all hits at $t = 50\%$ ZM bound.

TINS hits as negative allosteric modulators (NAMs) of A2aR

Fragment	k_{off} of [^3H]ZM241385 (min $^{-1}$) ^a
Control ZM241385	0.011 + 0.002
+ 2.5mM ZB418	0.040 0.007
+ 2.5mM ZB2044	0.023 0.006
+ 2.5mM ZB1153	0.016 0.005

^a $k_{\text{off}} \pm \text{SEM}$ (n=3).

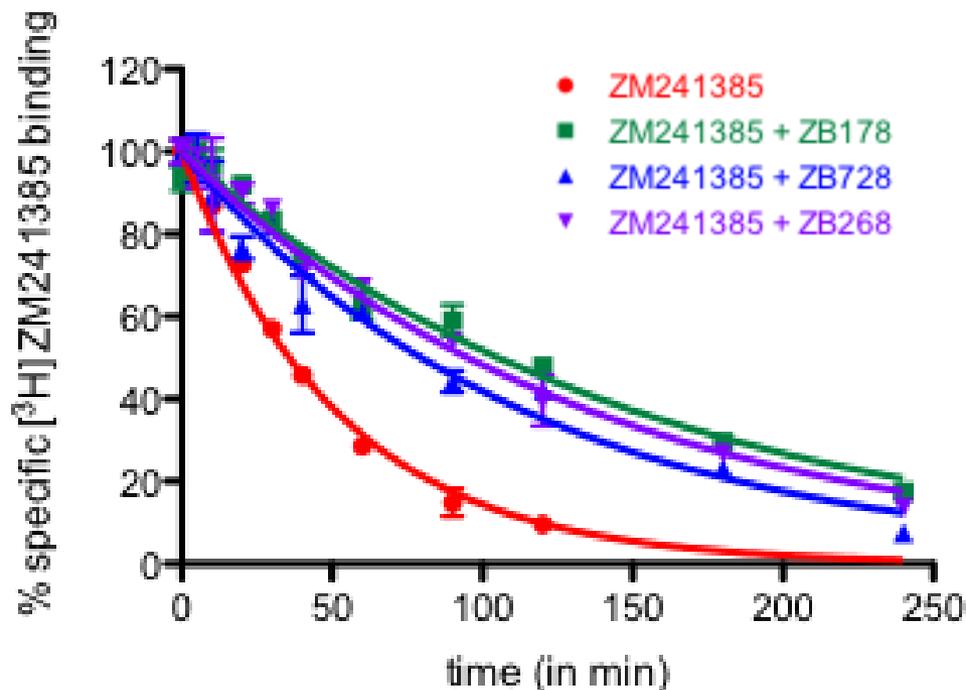
NAMs



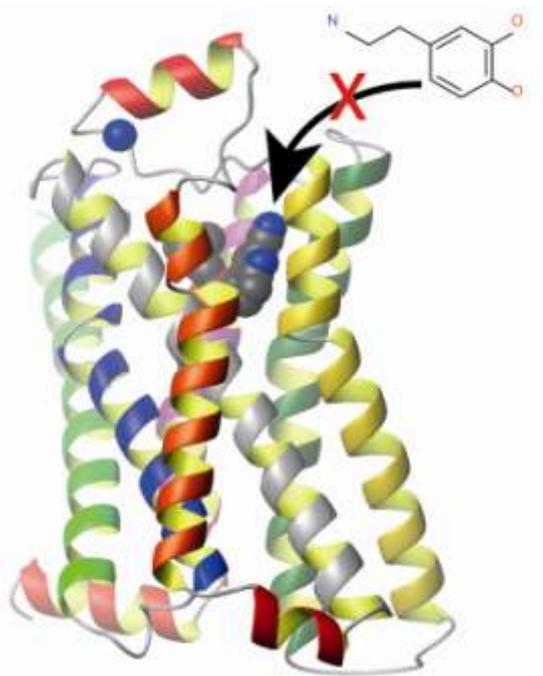
TINS hits as positive allosteric modulators (PAMs) of A2aR

Fragment	k_{off} of [^3H]ZM241385 (min $^{-1}$) ^a
Control ZM241385	0.0109 + 0.0018
+ 2.5mM ZB178	0.0057 0.0011
+ 2.5mM ZB728	0.0069 0.0018
+ 2.5mM ZB268	0.0066 0.0007

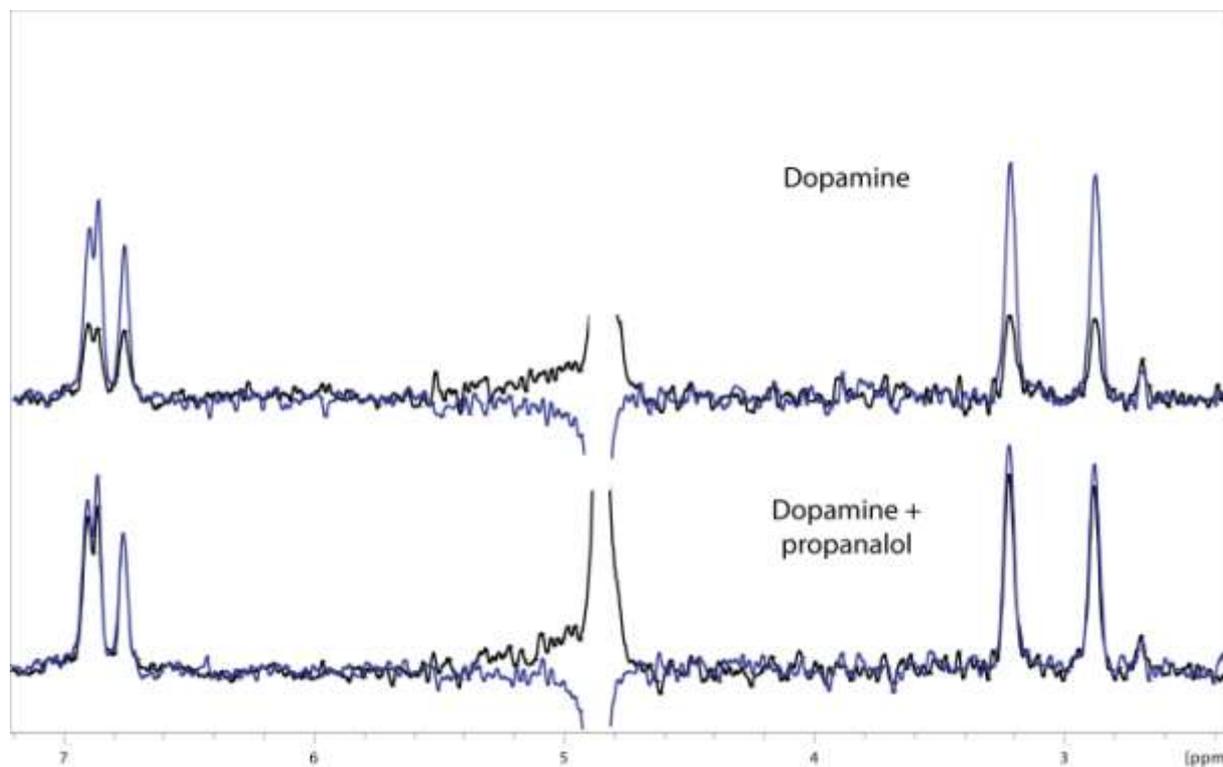
PAMs



Structural information from TINS?



β_1 AR



Publications

- Comparison of fragment screening methods – Kobayashi et al., J. Biomol. Screen, 2010, **15**, p.978
- Fragment screening of membrane proteins in detergents and nanodiscs – Früh et al., 2010, Chem & Biol., **17**, p.881

Acknowledgements

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